



Studying the Interactions of a Gemini Surfactant with DNA by Fluorescence

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Outline

- **Introduction**

- Gemini surfactants in gene delivery
- Study the associations of a pyrene labeled gemini surfactant by fluorescence
 - With and without DNA

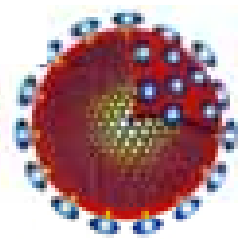
- **Results**

- Global analysis fits of the fluorescence decays of the pyrene monomer and excimer

- **Conclusions**

Gene Delivery

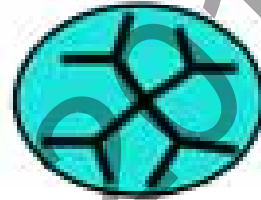
- **Types of delivery vectors**
 - **2 Classes: Viral and Non-Viral**



Viruses



Cationic Polymers



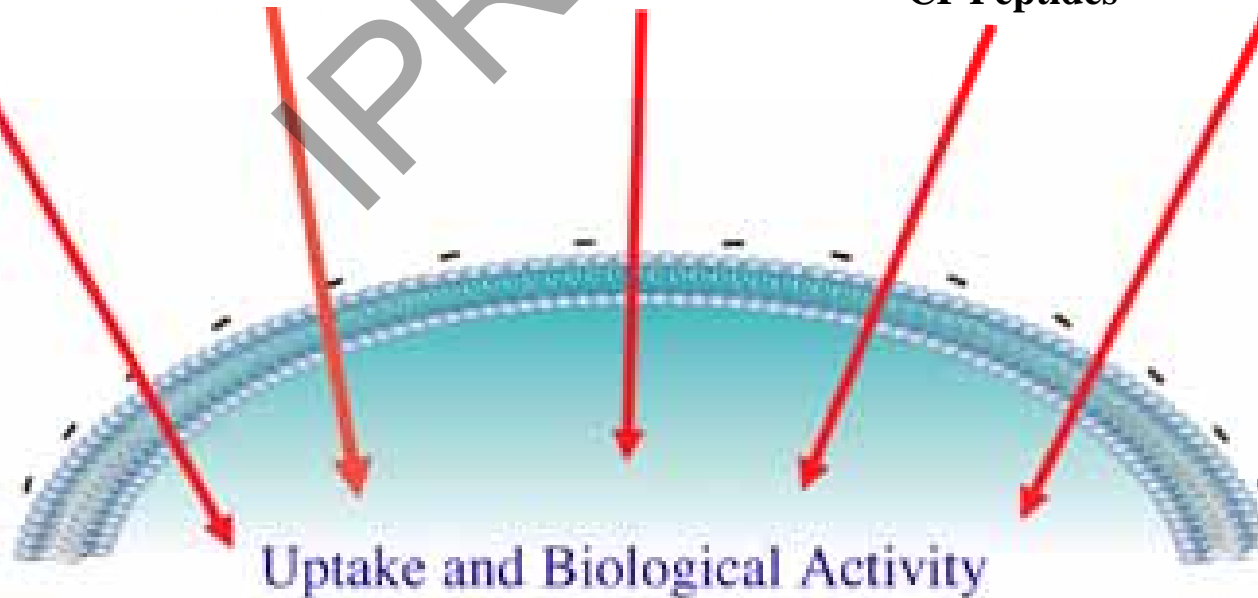
Dendrimers



CP Peptides



Liposomes



Gene Delivery Vectors

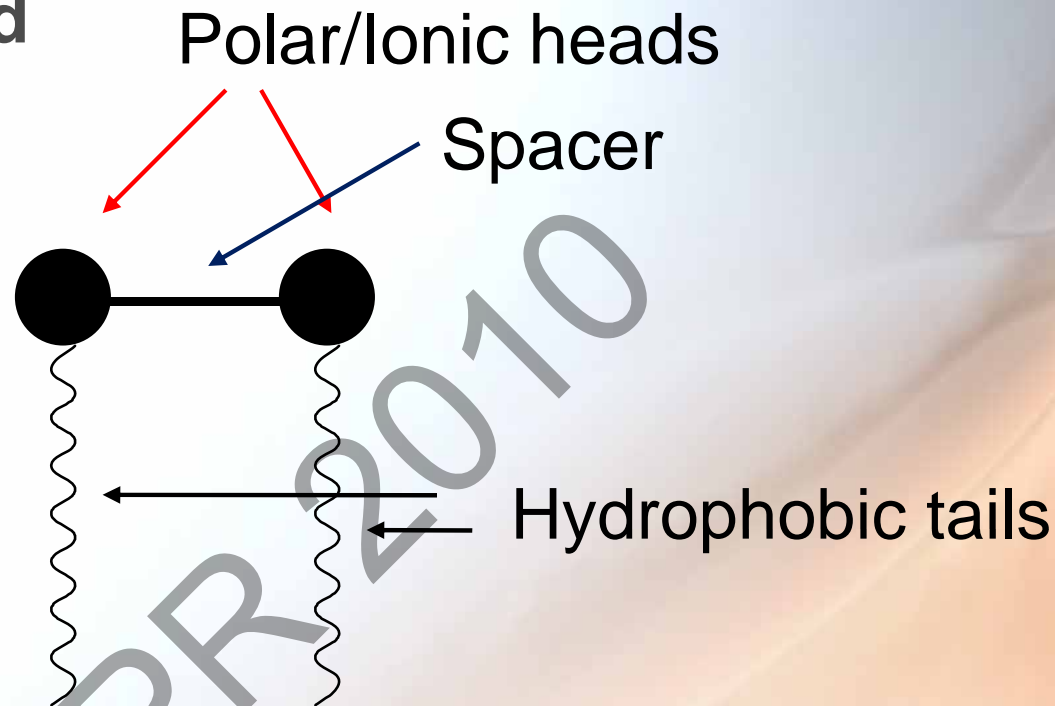
	VIRAL	NON-VIRAL
Advantages	<ul style="list-style-type: none">▪ Protects gene against degradation▪ High transfer efficiency	<ul style="list-style-type: none">▪ Non-toxic, non immunogenic▪ Not limited in the size of gene they can encapsulate▪ Relatively cheap
Disadvantages	<ul style="list-style-type: none">▪ Viral capsids have the potential to generate severe immune response▪ Limited in the size of the gene they can encapsulate	<ul style="list-style-type: none">▪ Low efficiency of transfection

Gao, X., Kim, K, Liu, D., *The AAPS Journal*, 9, 2007, 92 – 104.

Wettig, S. D., Verrall, R. E., Foldvari, M., *Current Gene Therapy*, 8, 2008, 9 – 23.

Gemini Surfactants

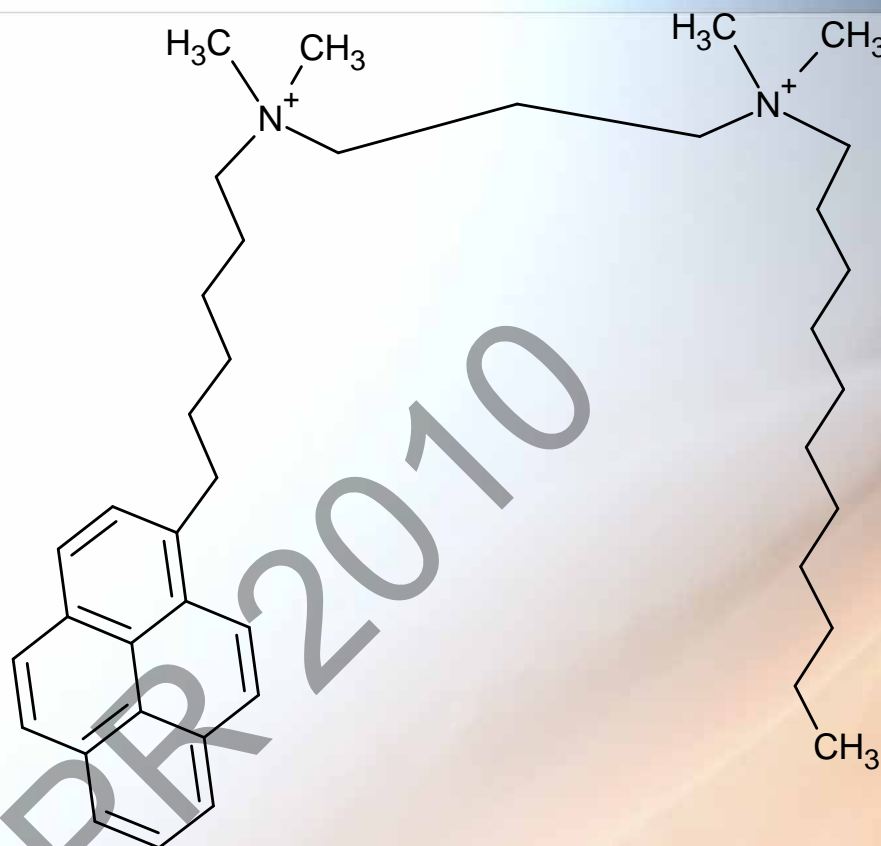
- **Contain 2 head and 2 tail groups that are chemically linked**



- **Spacer: short or long methylene groups, rigid (stilbene), polar (polyether), and non polar (aliphatic, aromatic).**
- **Gemini surfactants can also be asymmetrical.**

Pyrene Labeled Gemini Surfactant

[Py-3-12]

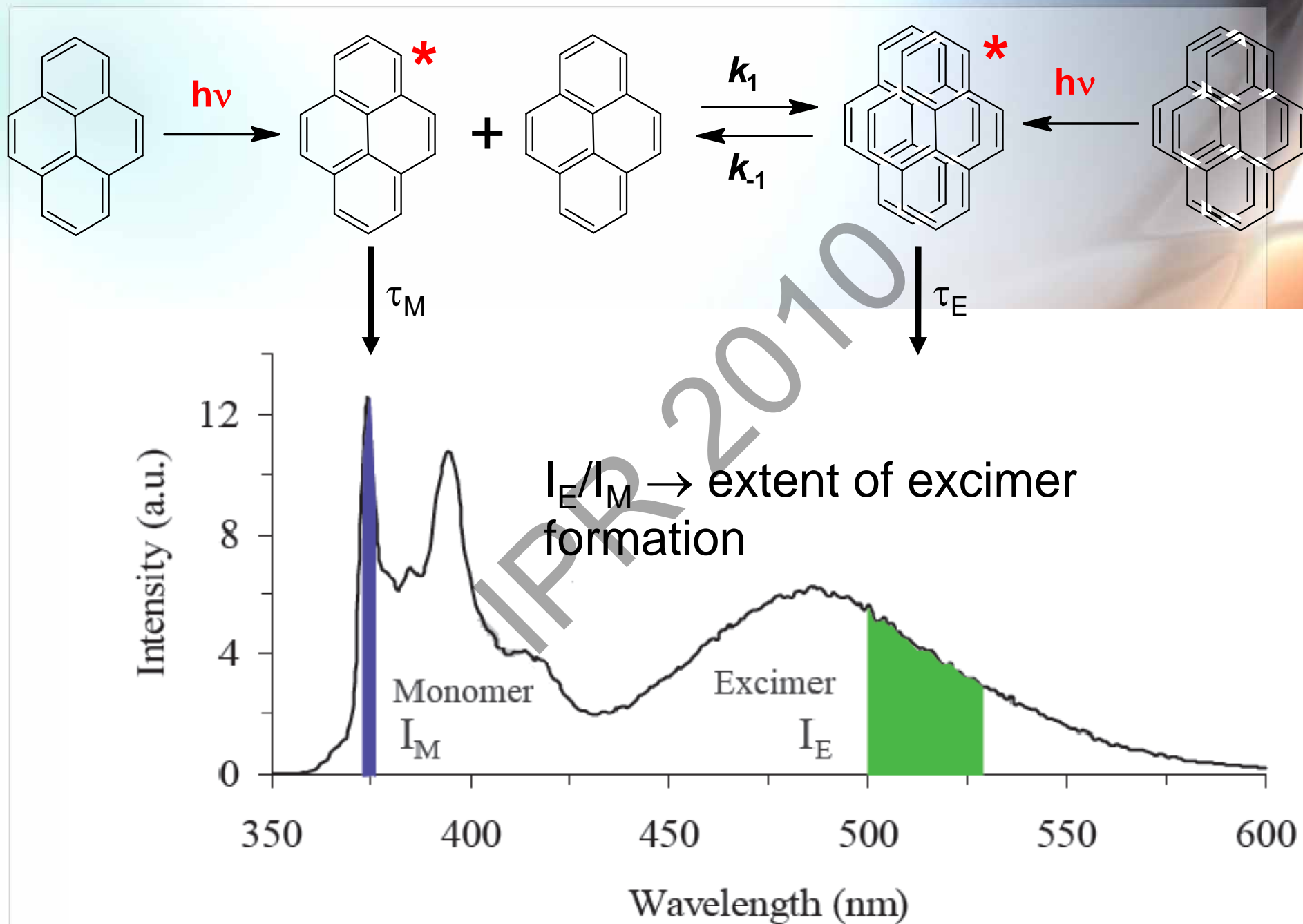


- **Gemini surfactant labeled with pyrene – study the gemini surfatcant complexes with DNA in solution by fluorescnece**
 - 1) **Characterize the micellization without DNA**
 - 2) **Characterize the complexes with DNA**
- **Given to us by Dr. Shawn Wettig from the Dept. of Pharmacy at UW – CMC = 0.22 mM**

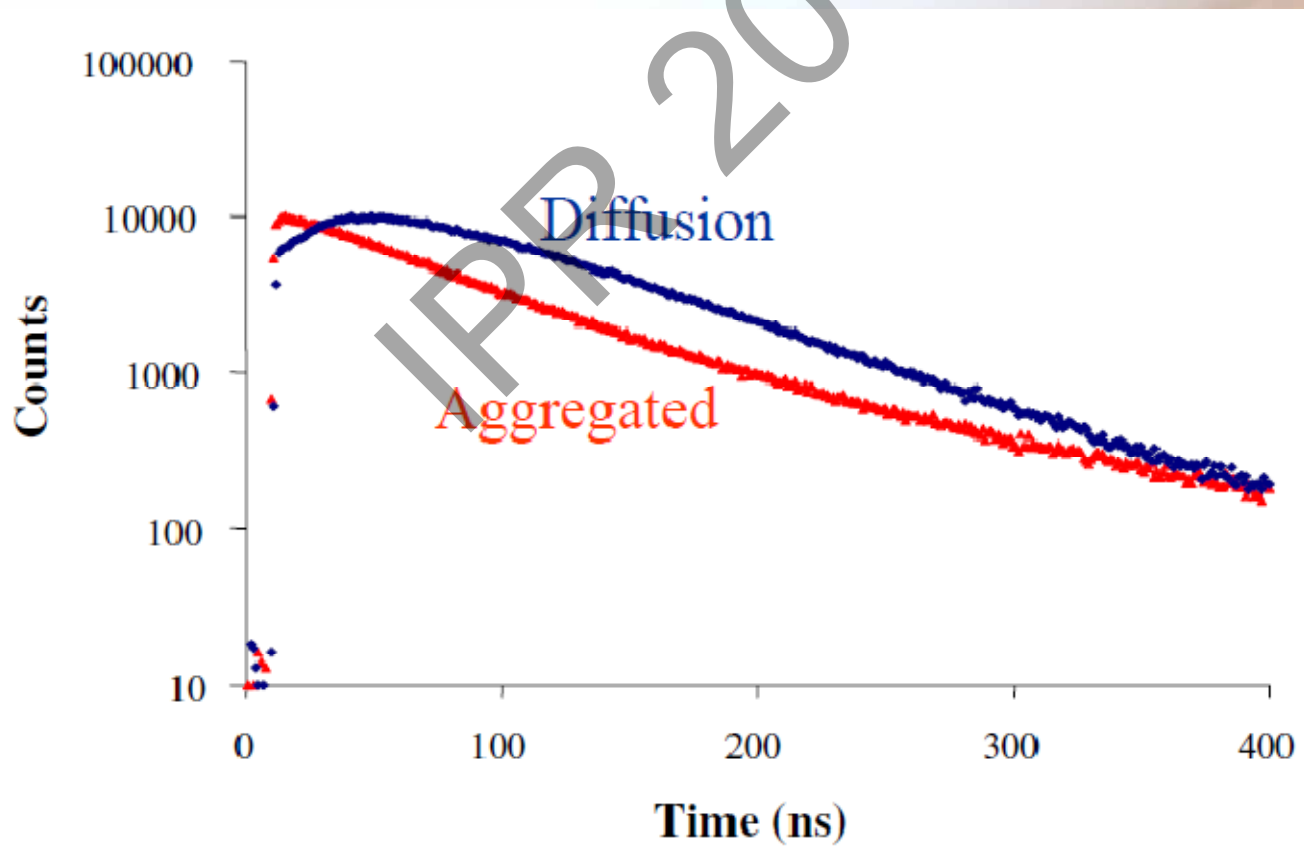
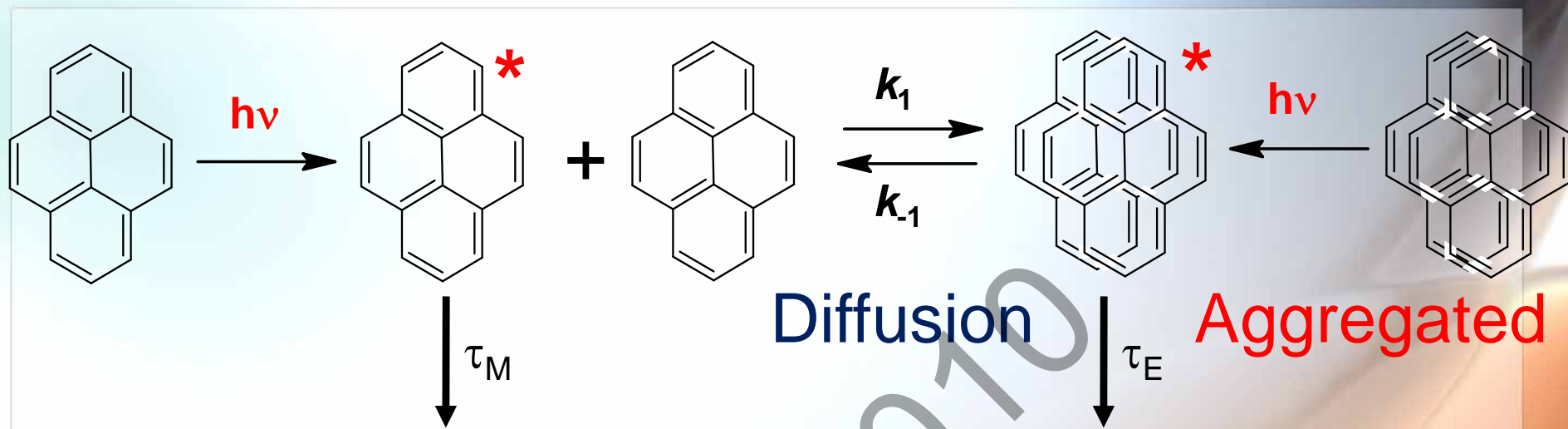
Why Pyrene?

- **Pyrene – has many interesting photophysical properties**
- **Pyrene's ability to form an excimer (excited dimer)**
- **Used to study the association of pyrene-labeled macromolecules in aqueous solution**

Pyrene - Fluorescence

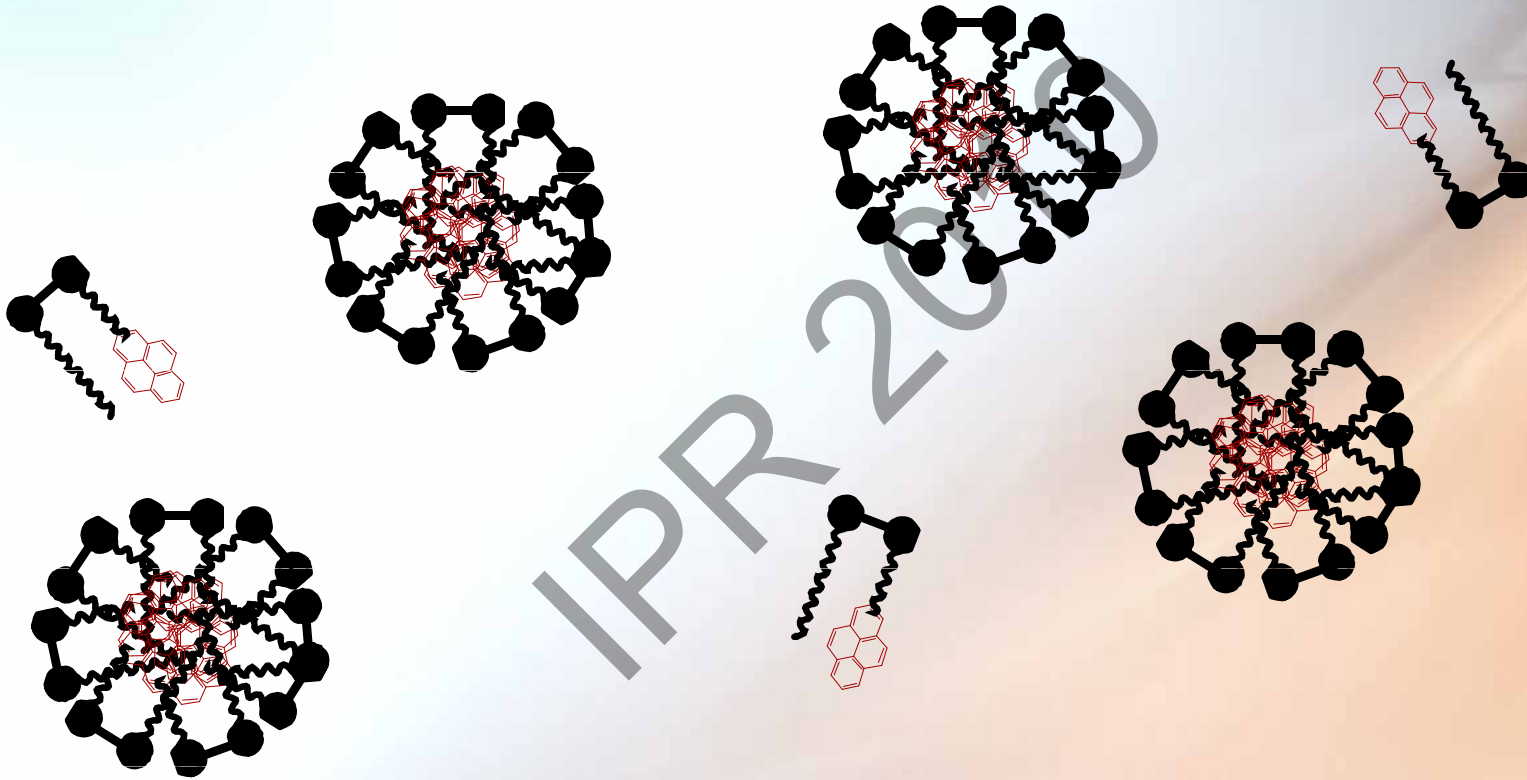


Pyrene - Excimer Decays



Pyrene Labeled Surfactant

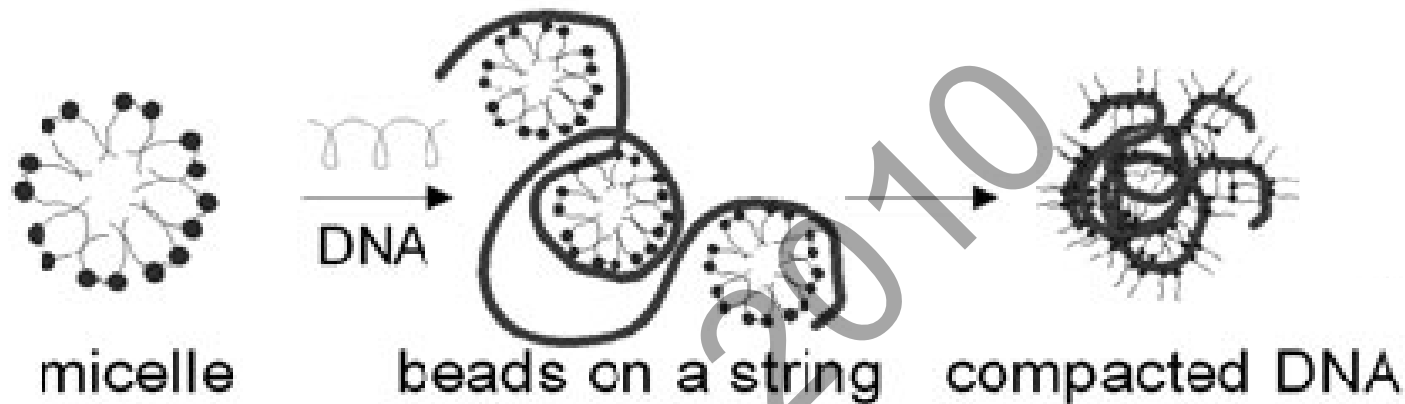
- **Without DNA**



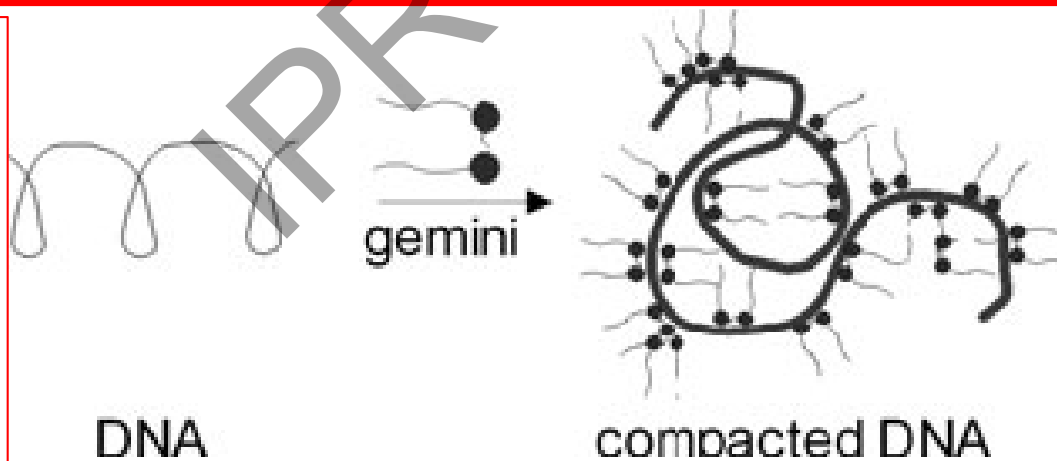
- **Fluorescence data gives us information about the CMC**

Pyrene Labeled Surfactant

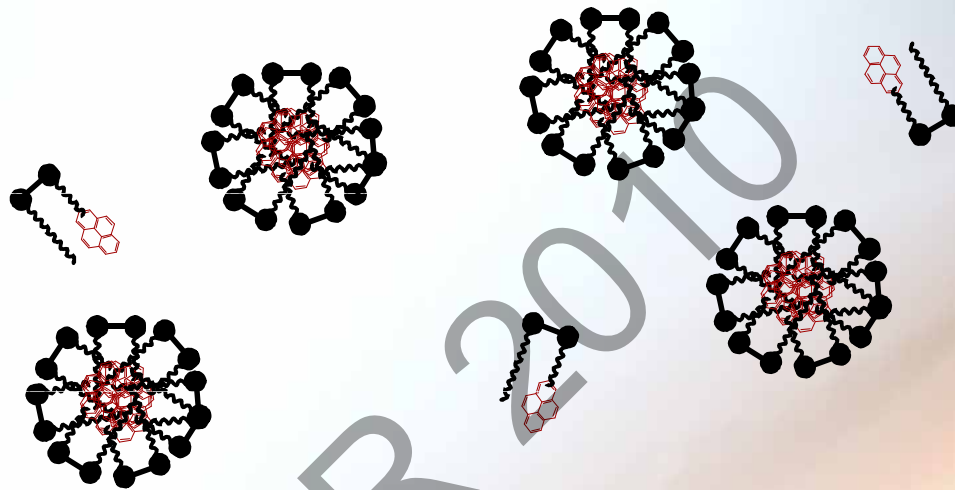
- **With DNA**



Excimer formation is due to complexation with DNA and not from micellization of the gemini surfactant



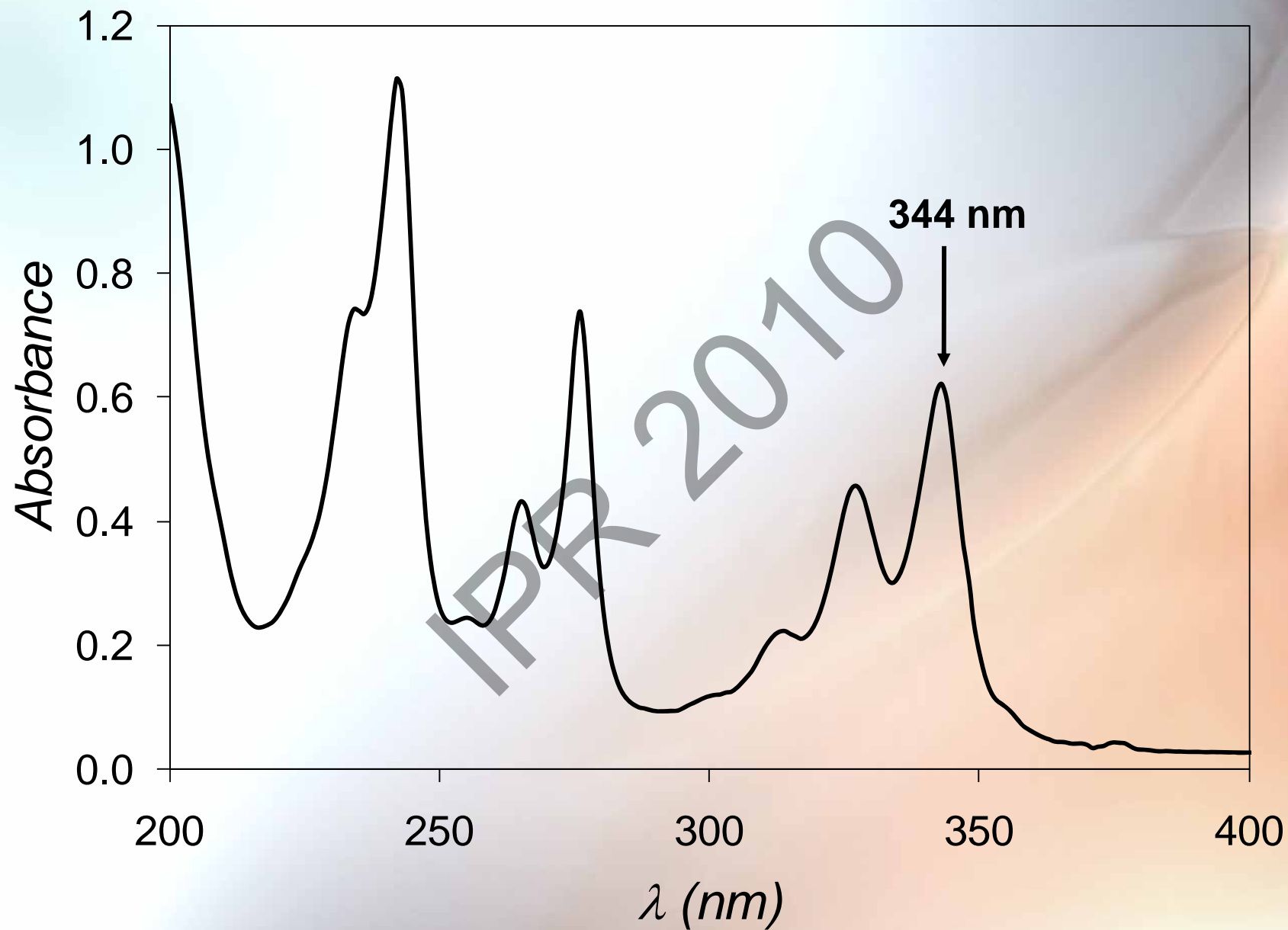
Based on AFM results



RESULTS

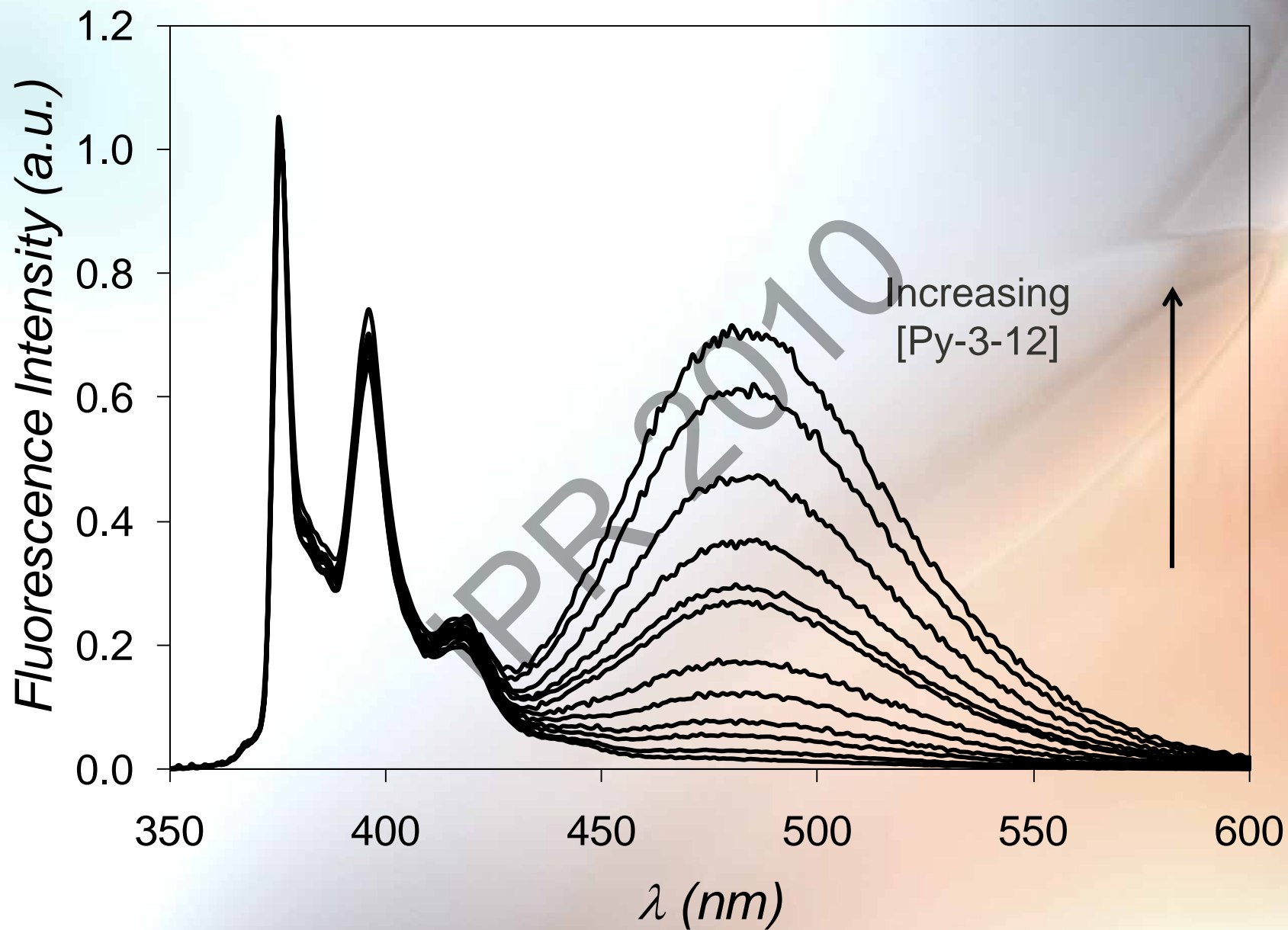
Micellization of Py-3-12 Without DNA

Absorbance Spectrum



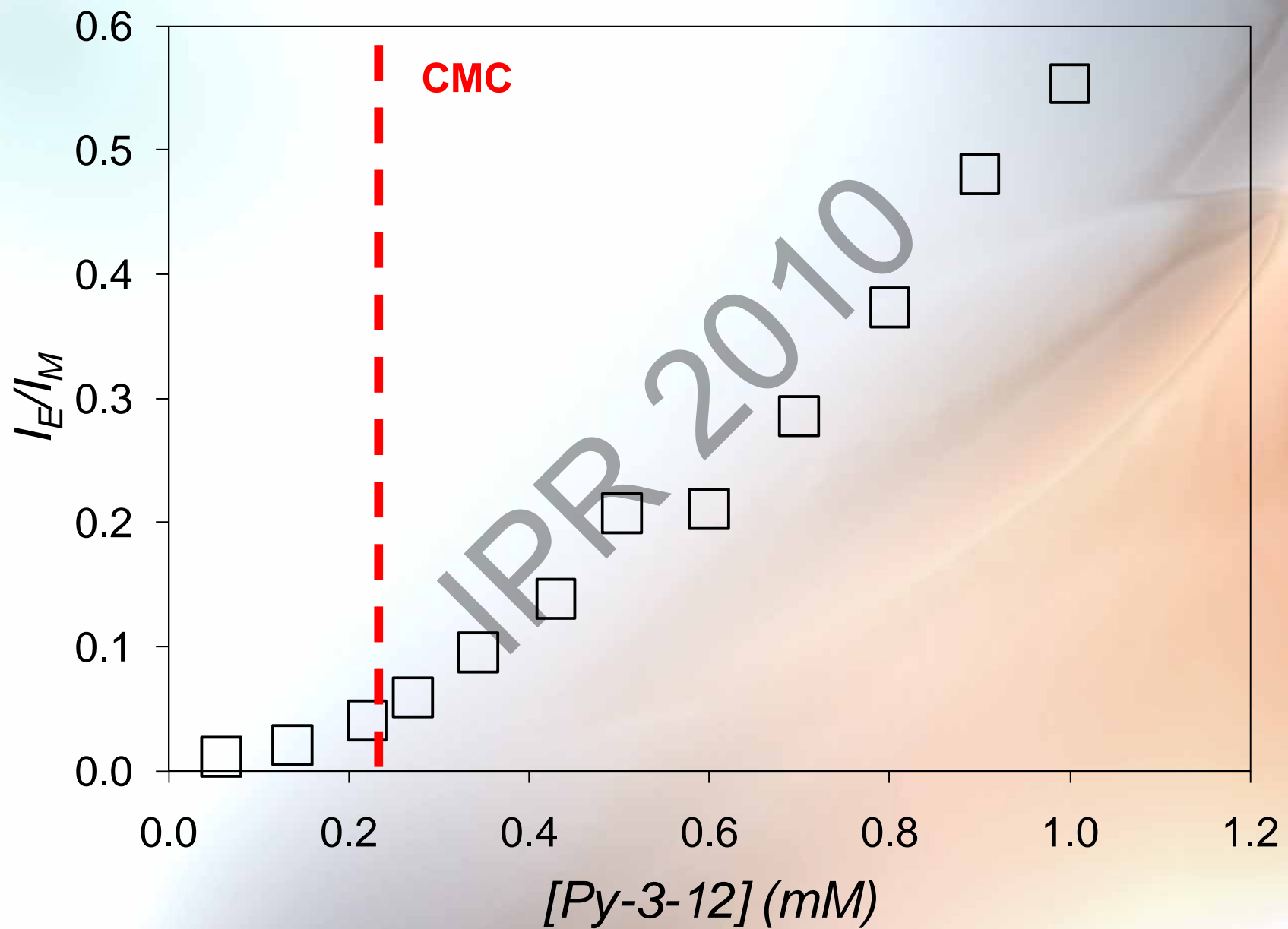
[Py-3-12] = 0.02 mM

Steady State Fluorescence

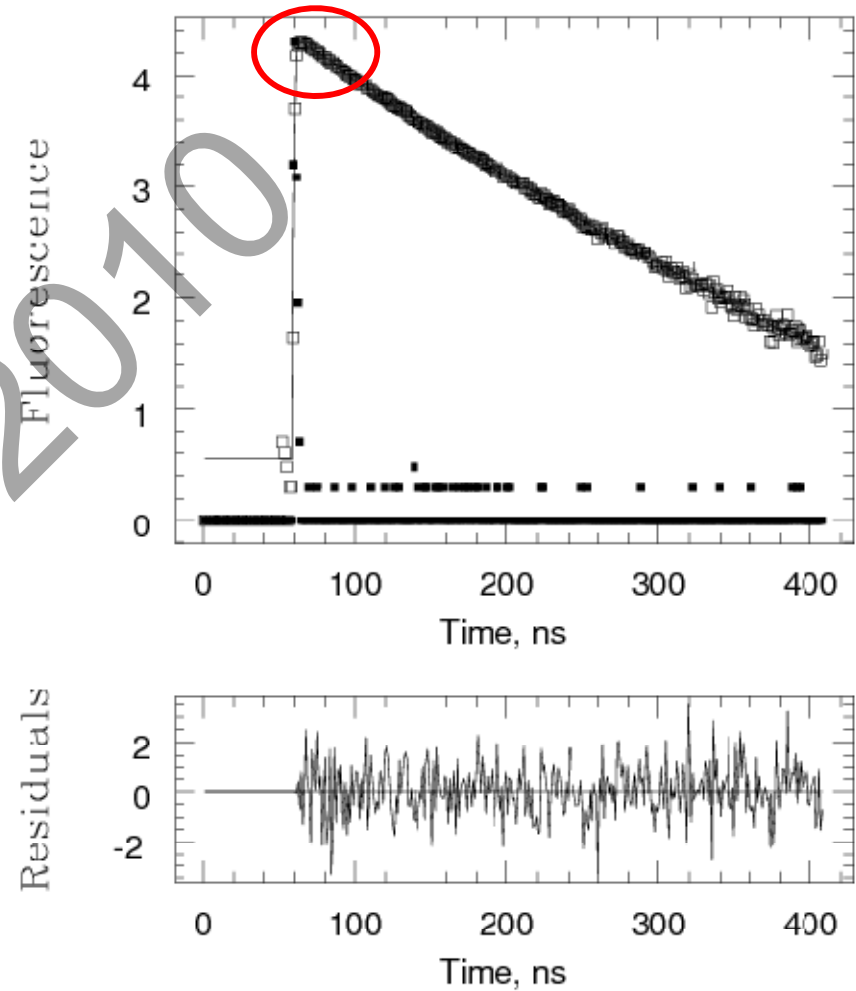
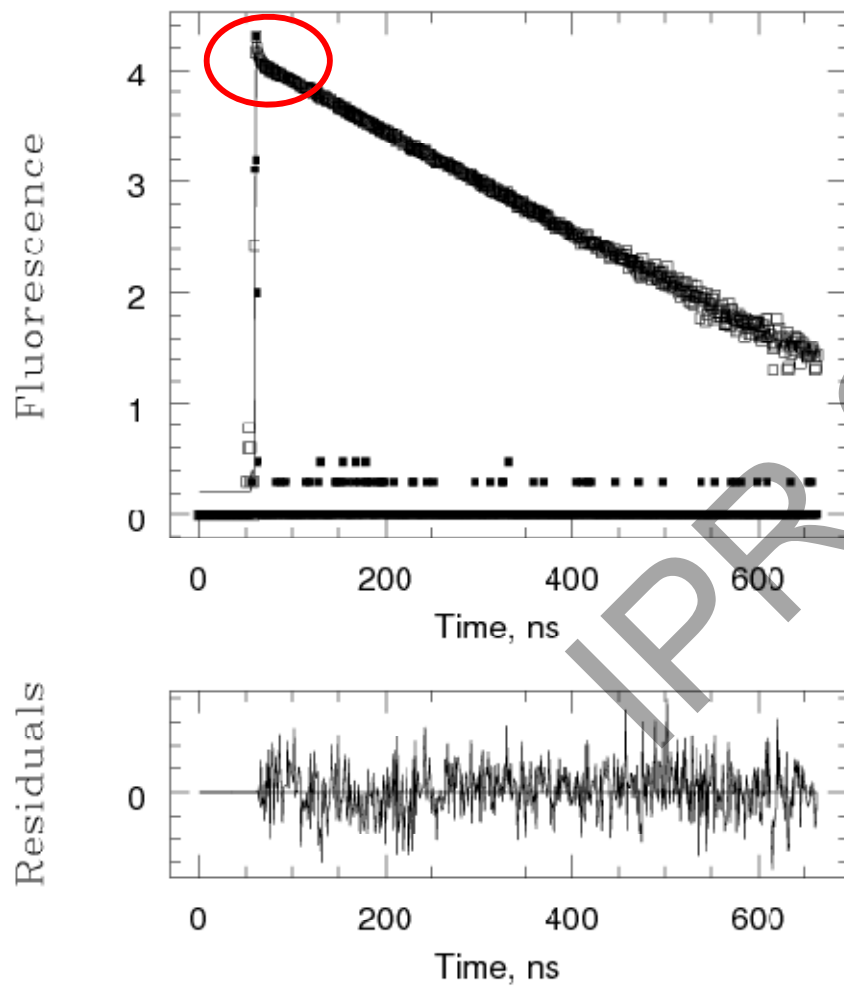


Normalized @ 375 nm

Steady State Fluorescence



Time Resolved Fluorescence Decays

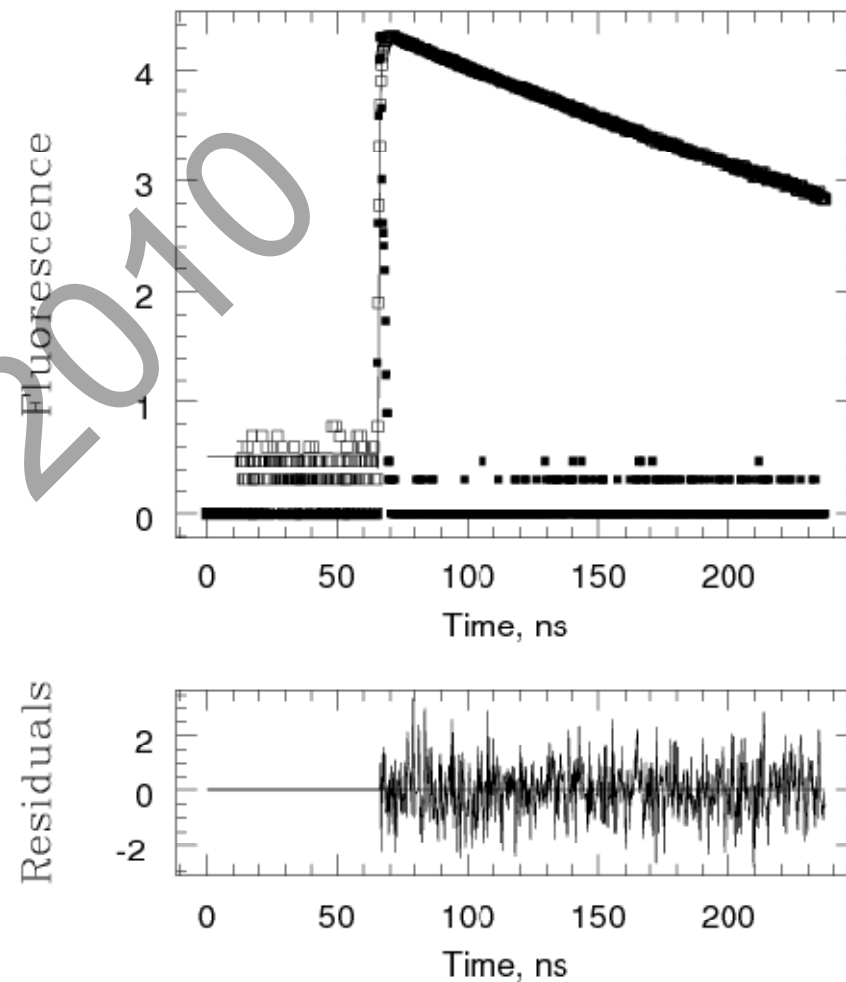
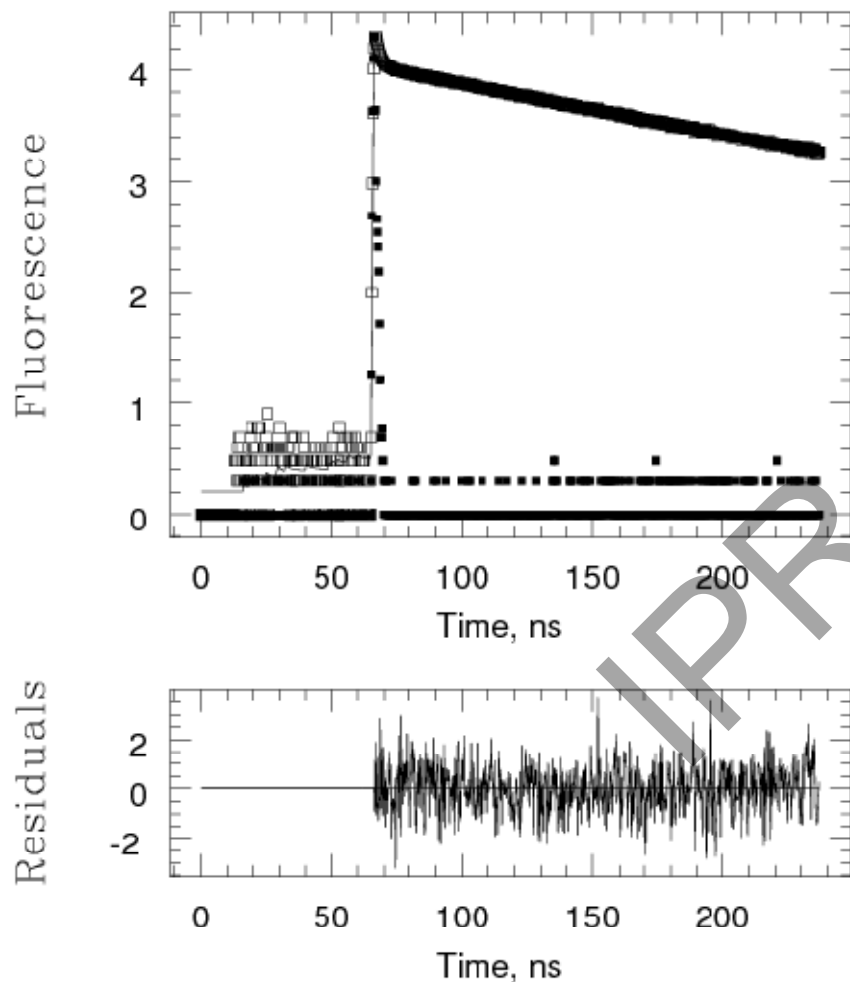


Time per Channel = 1.02 ns /channel → equivalent to short decay time

Time Resolved Fluorescence

Decays

Time per Channel = 0.24 ns /channel



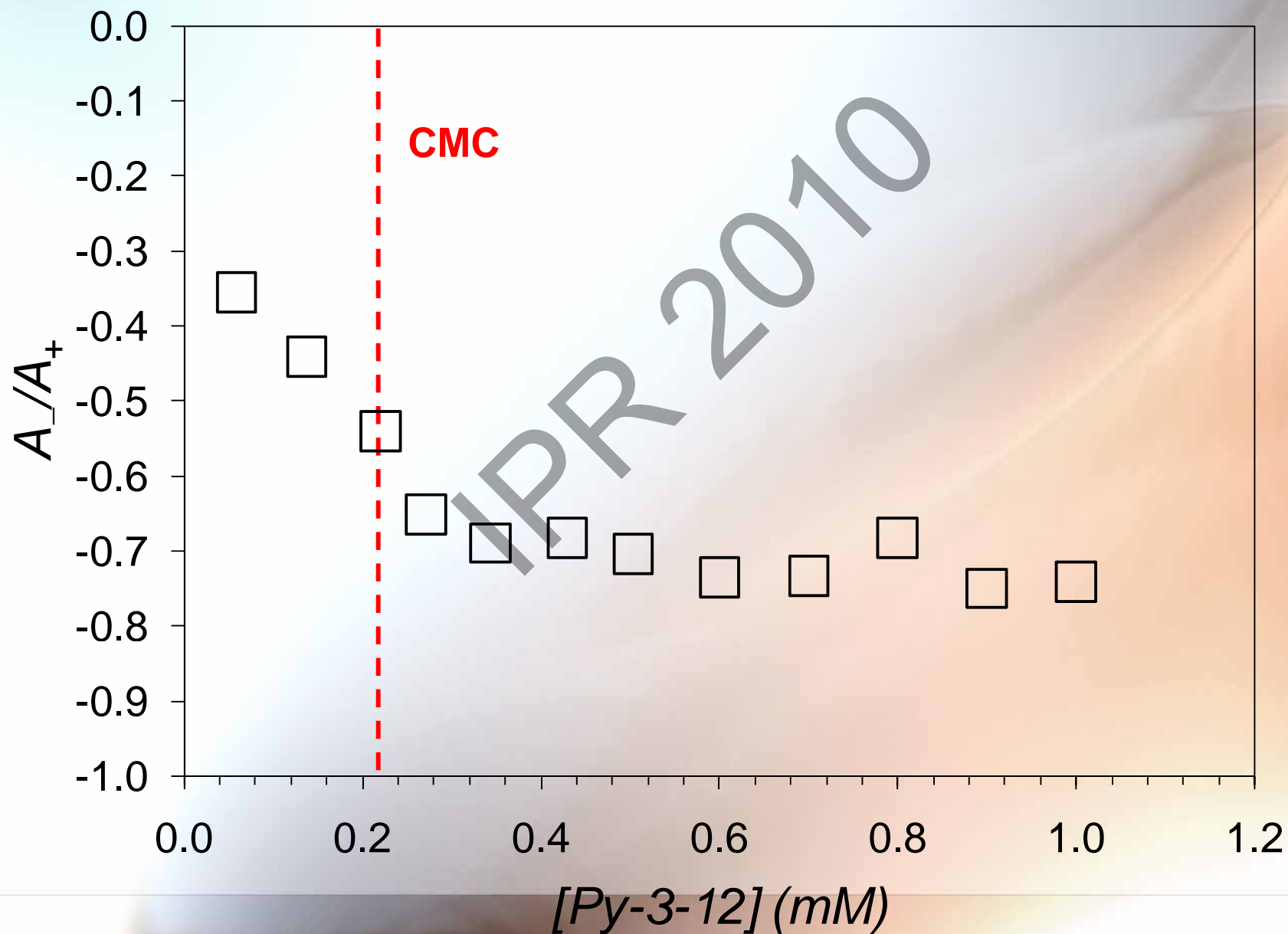
$$I(t) = A_1 e^{-t/\tau_1} + A_2 e^{-t/\tau_2} + A_3 e^{-t/\tau_3}$$

$A_-/A_+ \rightarrow -1$ excimer formed via diffusion $a_-/a_+ \rightarrow 0$ ground state dimers

Time Resolved Fluorescence

Decays

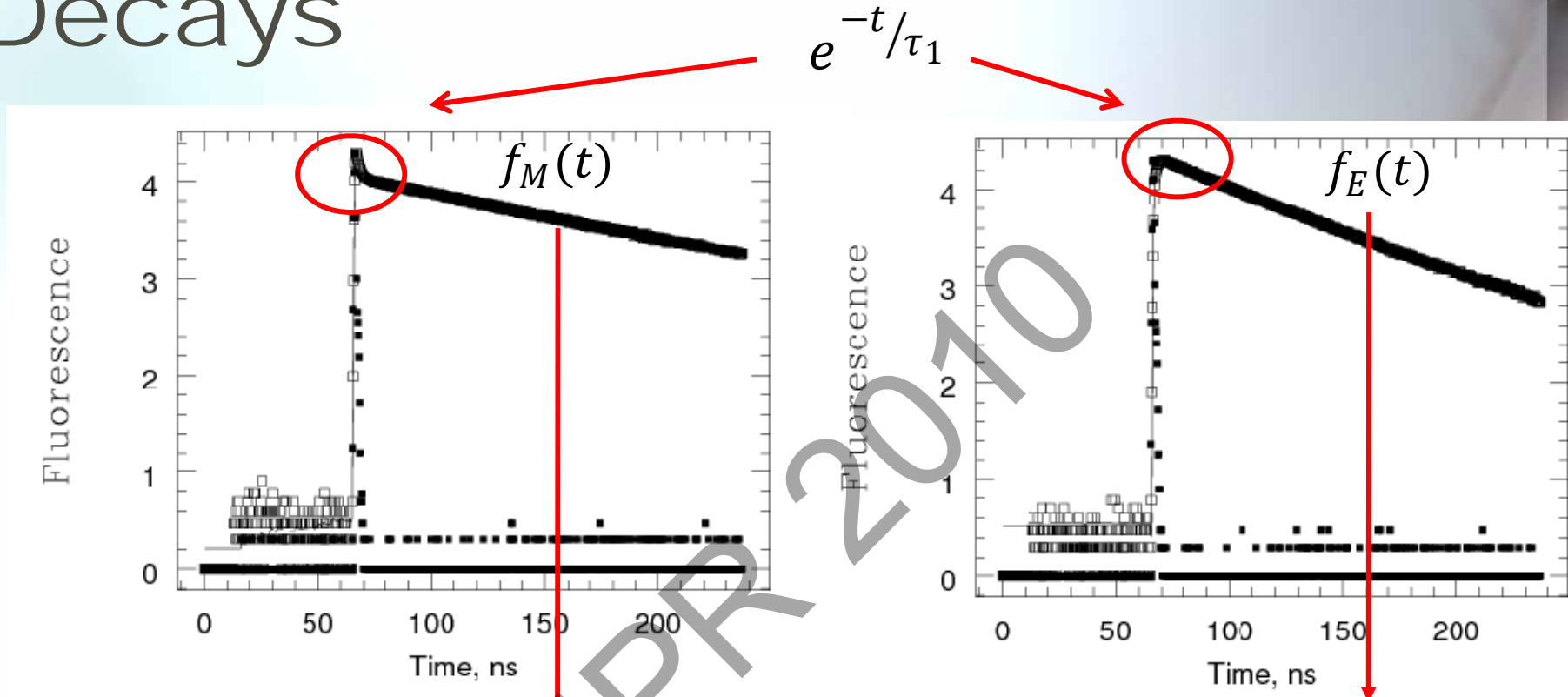
$A_-/A_+ \rightarrow -1$ excimer formed via diffusion $A_-/A_+ \rightarrow 0$ ground state dimers



Global Analysis

- **Excimer formation via diffusion**
 - Kinetics of monomer consumption and excimer formation are coupled
 - Decay times in the monomer and excimer (rise time) are found to be the same when fit with a sum of exponentials
 - Fit the monomer and excimer globally by ensuring some of the decay times are held the same in the monomer and excimer decays
 - Increases the accuracy on the parameters retrieved from the fluorescence decays

Time Resolved Fluorescence Decays



$$a_1 e^{-t/\tau_{M1}} + a_2 e^{-t/\tau_{M2}}$$

$$a_{E0} e^{-t/\tau_{E0}} + a_D e^{-t/\tau_D}$$

Lifetime of pyrene monomer ~ 97 ns

pyrenes that form excimer (proper stacking)

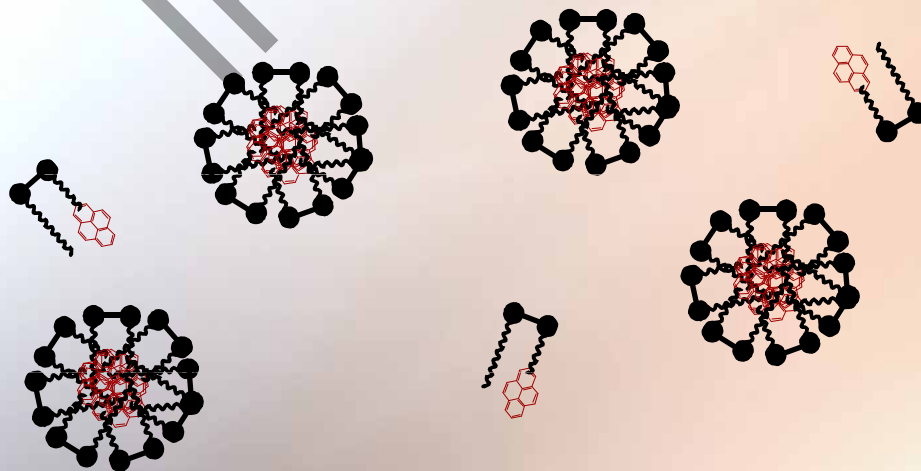
Impurity ~ 20 - 6 ns (3%)

pyrenes that form long lived excimer (improper stacking)

Time per Channel = 0.24 ns /channel

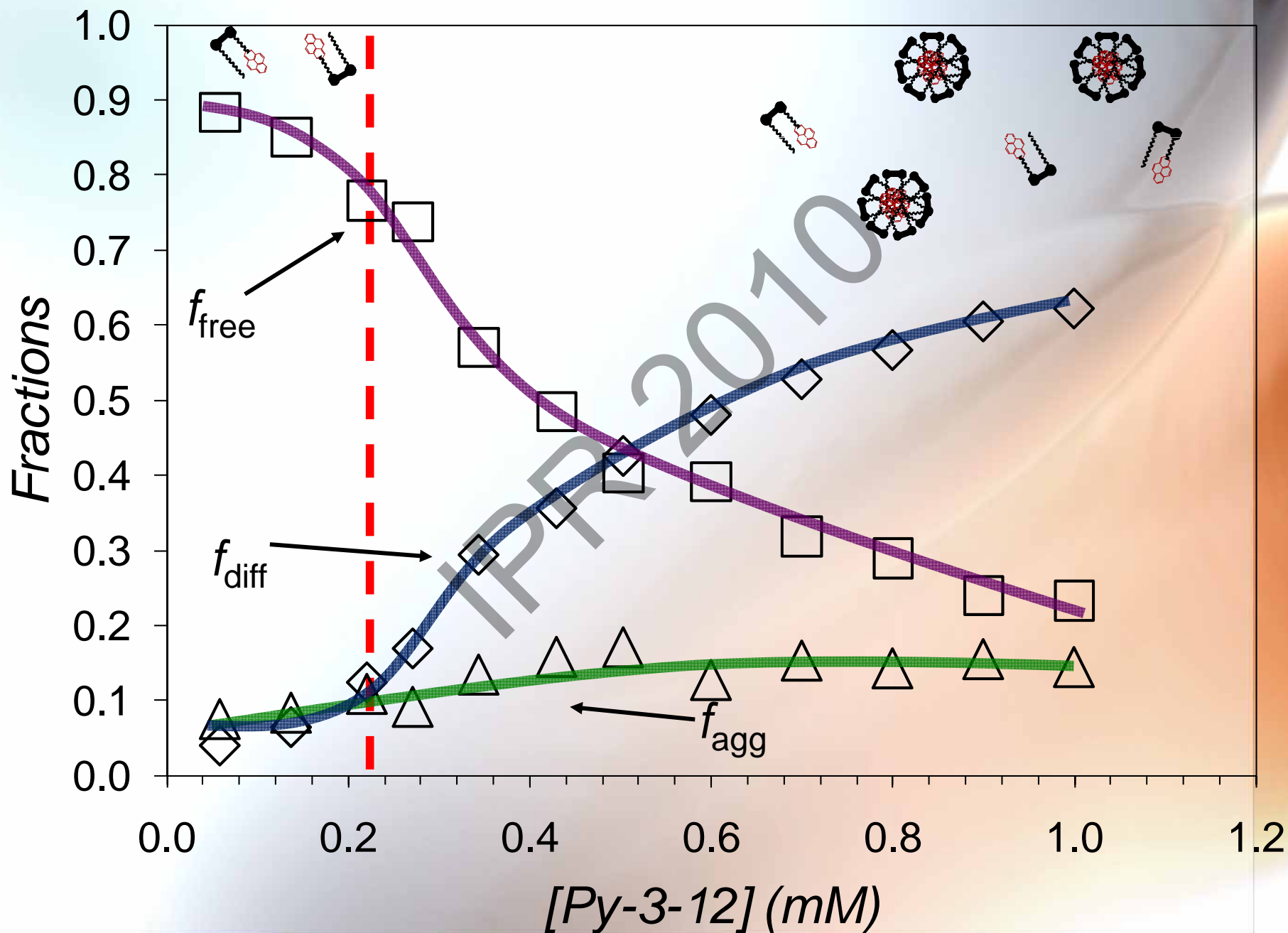
Global Analysis

- f_{free} – fraction of free pyrene
- f_{diff} – fraction pyrene that forms excimer via diffusion
- f_{E0} – fraction of preassociated pyrenes that form excimer (proper stacking)
- f_{D} – fraction of preassociated pyrenes that form long lived excimer (improper stacking)
- $f_{\text{agg}} = f_{\text{E0}} + f_{\text{D}}$

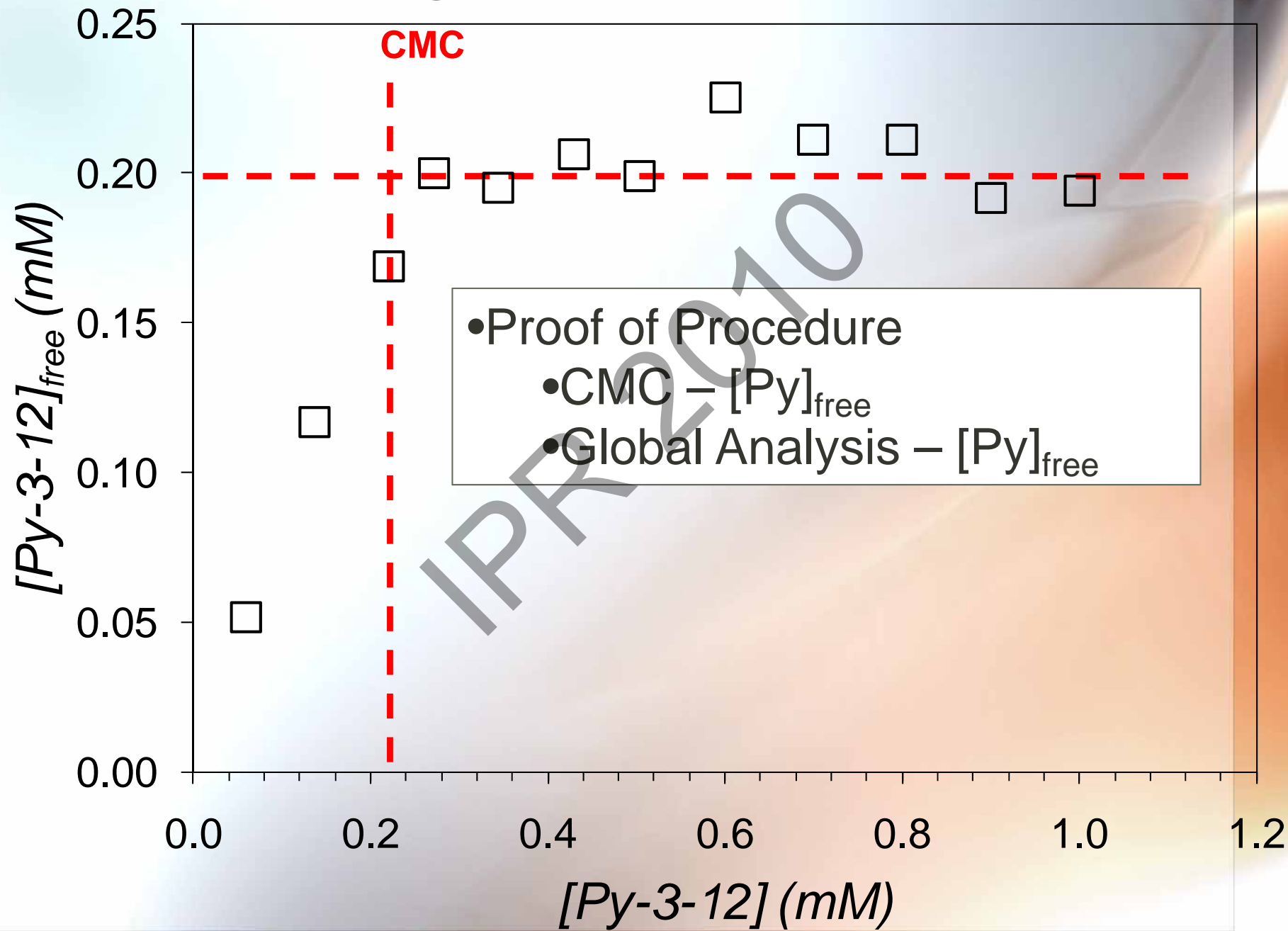


Global Analysis

CMC



Global Analysis



Global Analysis - Calculation

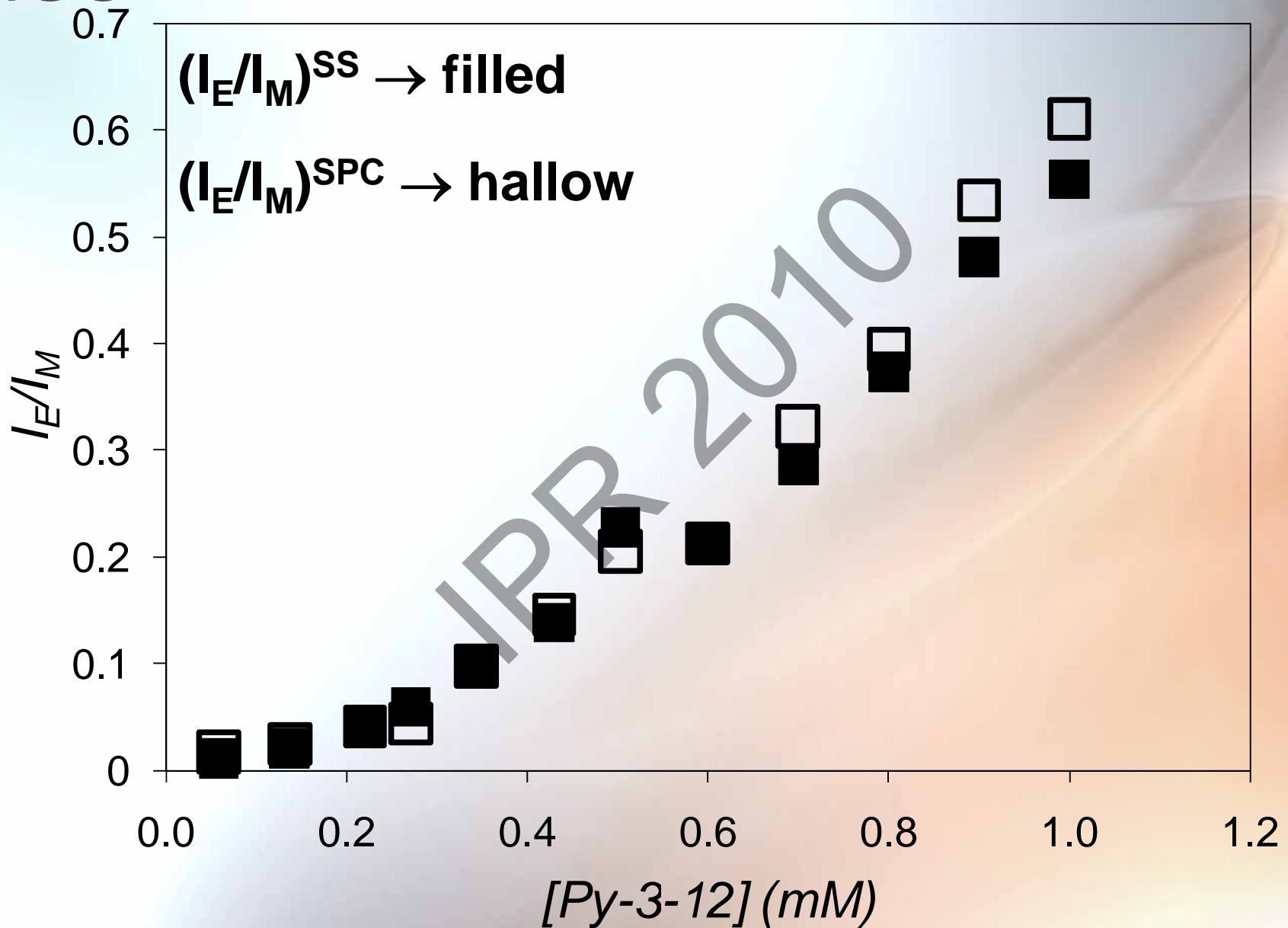
of $(I_E/I_M)^{SPC}$

$$I_M = \int_0^{\infty} [Py^*]_t dt = [Py]_o \left[f_{diff} \sum_{i=1}^n a_i \tau_i + f_{free} \tau_M \right]$$

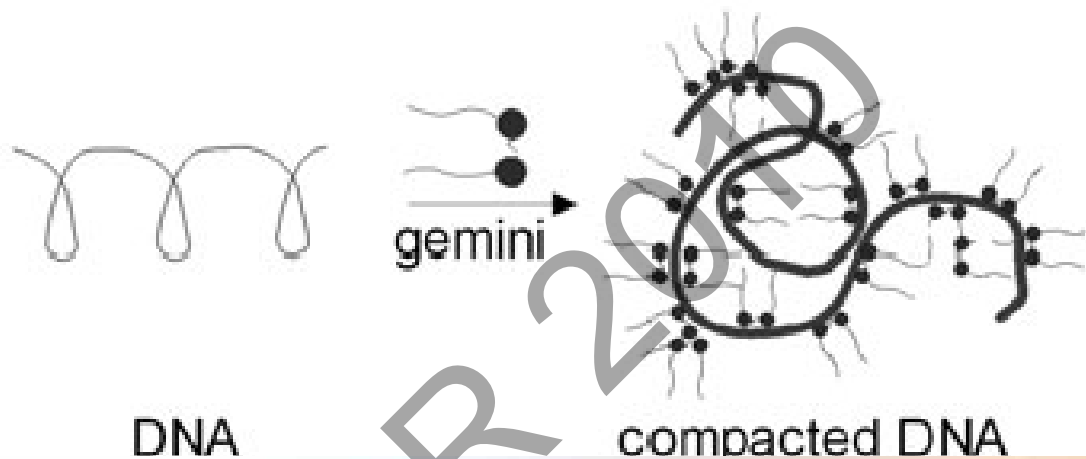
$$I_E = \int_0^{\infty} [E^*]_t dt$$

$$= [Py]_o \left[-f_{diffE0} \sum_{i=1}^n a_i \frac{\frac{1}{\tau_i} - \frac{1}{\tau_M}}{\frac{1}{\tau_i} - \frac{1}{\tau_{E0}}} \tau_i + \left(f_{E0} + f_{diffE0} \sum_{i=1}^n a_i \frac{\frac{1}{\tau_i} - \frac{1}{\tau_M}}{\frac{1}{\tau_i} - \frac{1}{\tau_{E0}}} \right) \tau_{E0} \right. \\ \left. - f_{diffD} \sum_{i=1}^n a_i \frac{\frac{1}{\tau_i} - \frac{1}{\tau_M}}{\frac{1}{\tau_i} - \frac{1}{\tau_D}} \tau_i + \left(f_D + f_{diffD} \sum_{i=1}^n a_i \frac{\frac{1}{\tau_i} - \frac{1}{\tau_M}}{\frac{1}{\tau_i} - \frac{1}{\tau_D}} \right) \tau_D \right]$$

$(I_E/I_M)^{SS}$ versus $(I_E/I_M)^{SPC}$; front-face



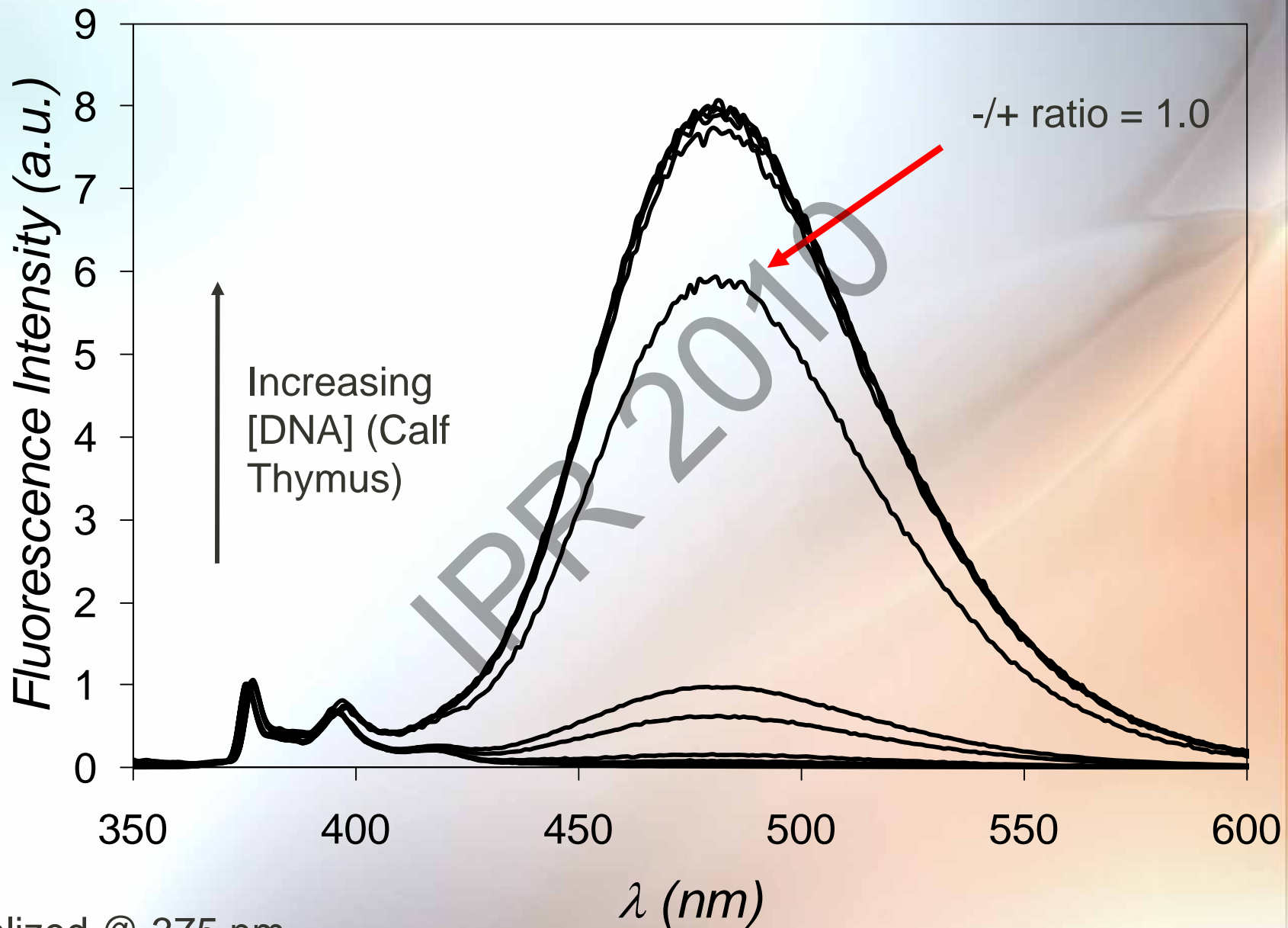
SPC trend scaled to SS trend



RESULTS

Complexation of Py-3-12 With DNA

Steady State Fluorescence

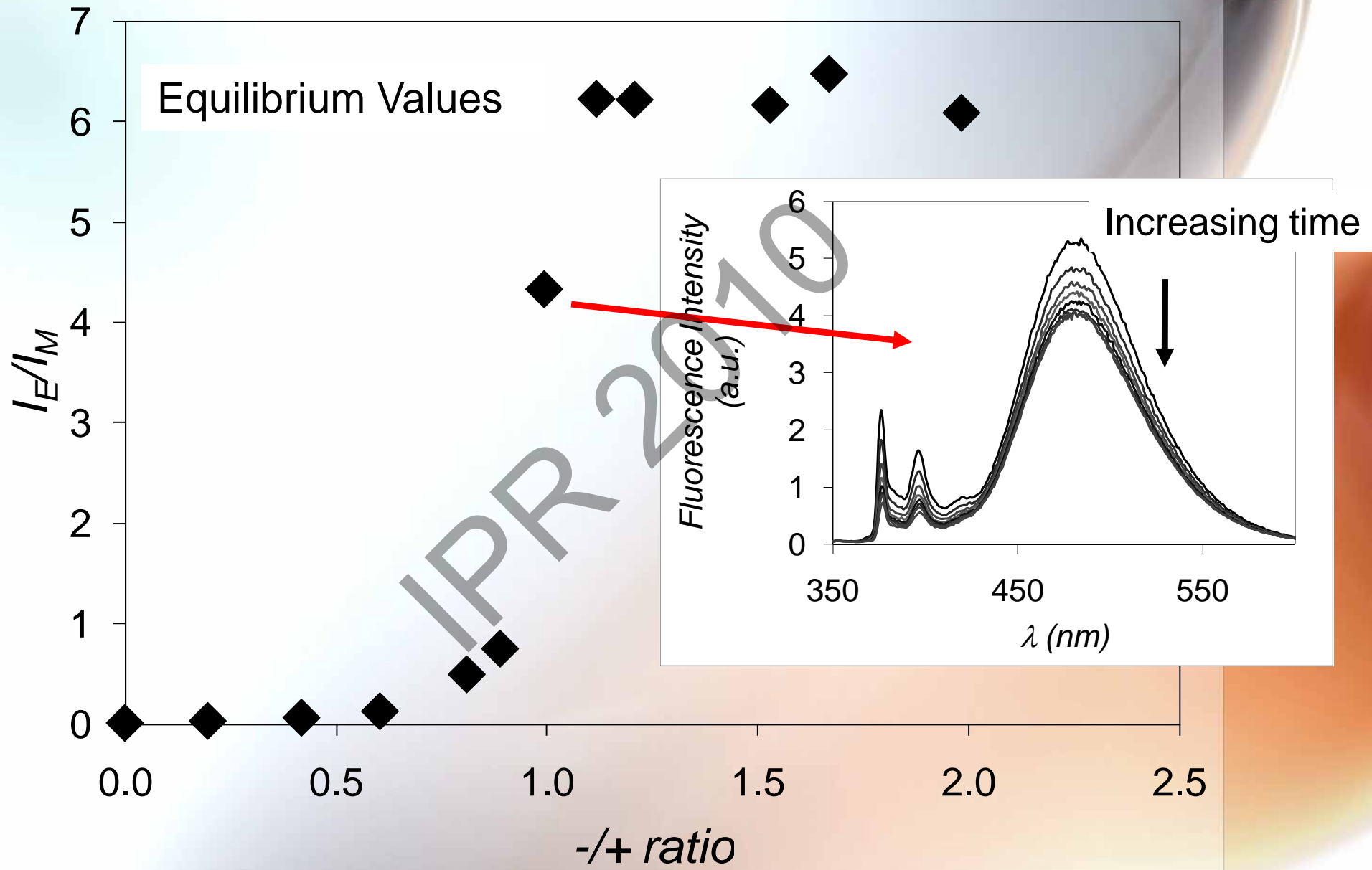


Normalized @ 375 nm

[Py-3-12] = 1×10^{-4} M

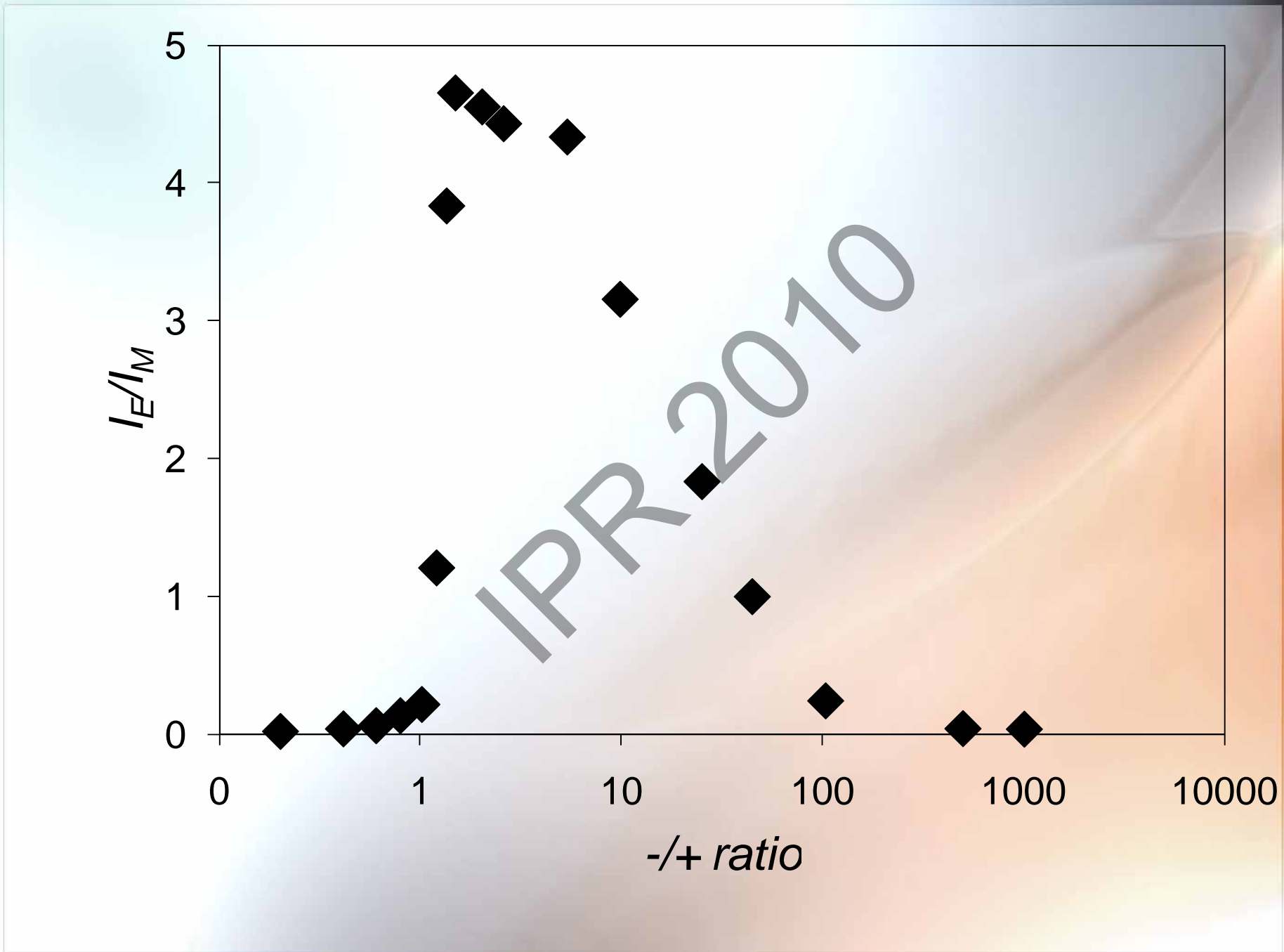
-/+ ratio \rightarrow [DNA-bp]/[Py-3-12]

Steady State Fluorescence



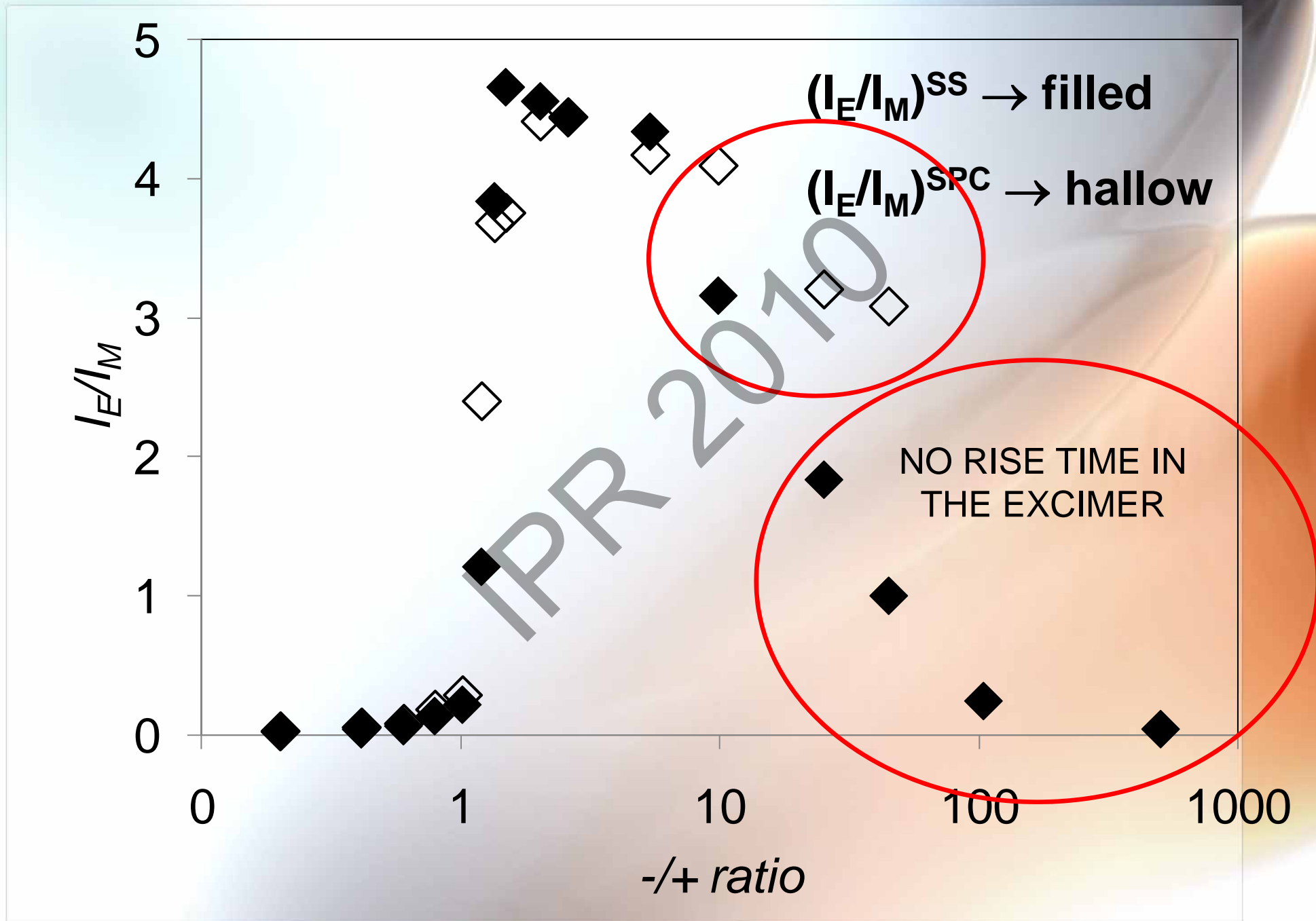
- Can we add enough DNA separate the surfactant molecules?
- $I_E/I_M \rightarrow 0$?

Steady State Fluorescence



Sheared Calf Thymus DNA – reduce viscosity at high $-/+$ ratios

$(I_E/I_M)^{SS}$ versus $(I_E/I_M)^{SPC}$



SPC trend scaled to SS trend

Global Analysis

