

Combination of SPME as non-invasive sample preparation technique and GCxGC-TOFMS for high resolution profiling of metabolites in apples: method development considerations and potential of new *invivo* SPME formats



Sanja Risticvic¹, Jennifer R. DeEll², Janusz Pawliszyn¹

¹ Department of Chemistry, University of Waterloo, Waterloo, Canada;
² Ontario Ministry of Agriculture, Food and Rural Affairs, Simcoe, Canada

Introduction

Results & Discussion

Solid phase microextraction: SPME

developed by Pawliszyn et al. in 1989

↓ disadvantages of traditional sample preparation;

combination of sampling, extraction, concentration & sample introduction into 1 solvent free step;

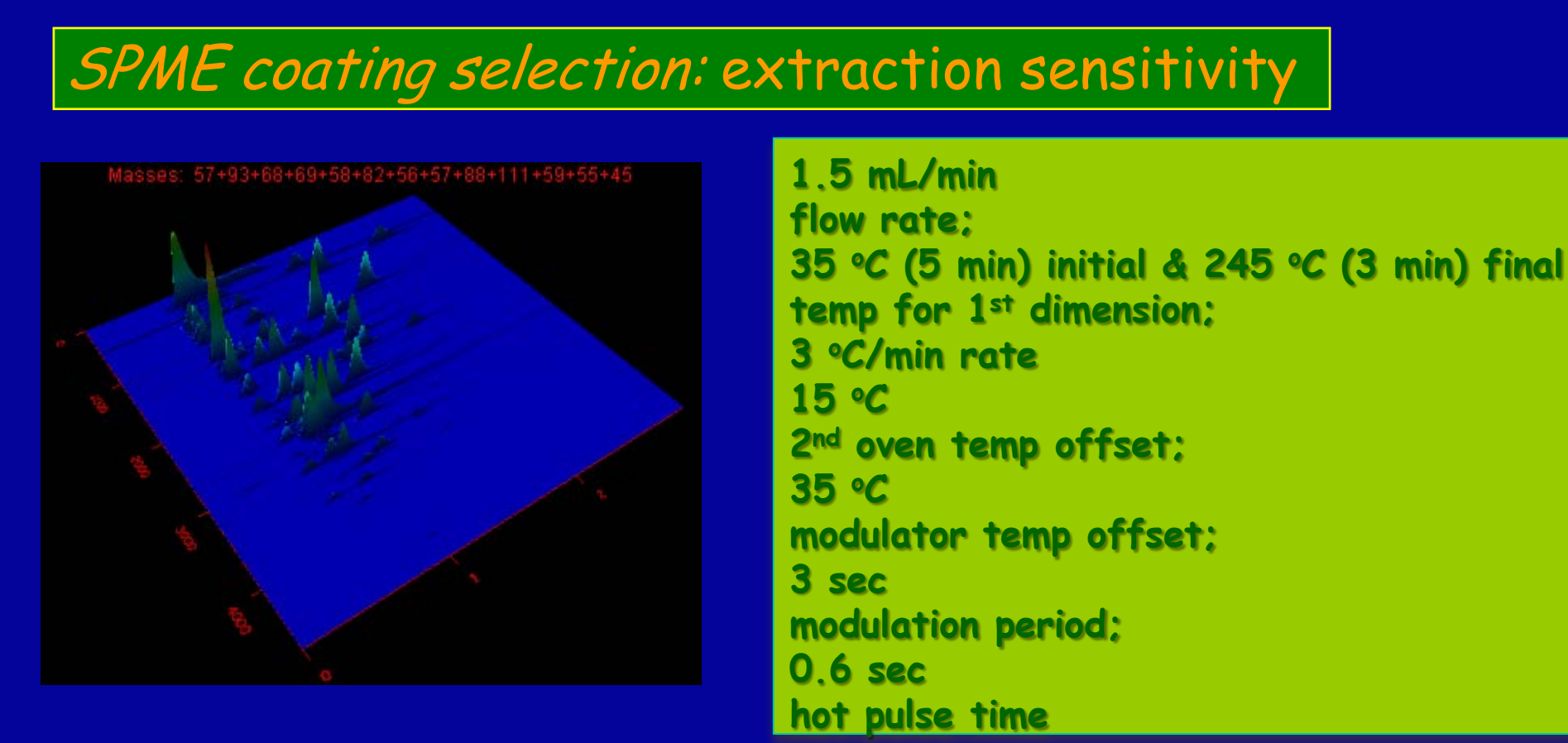
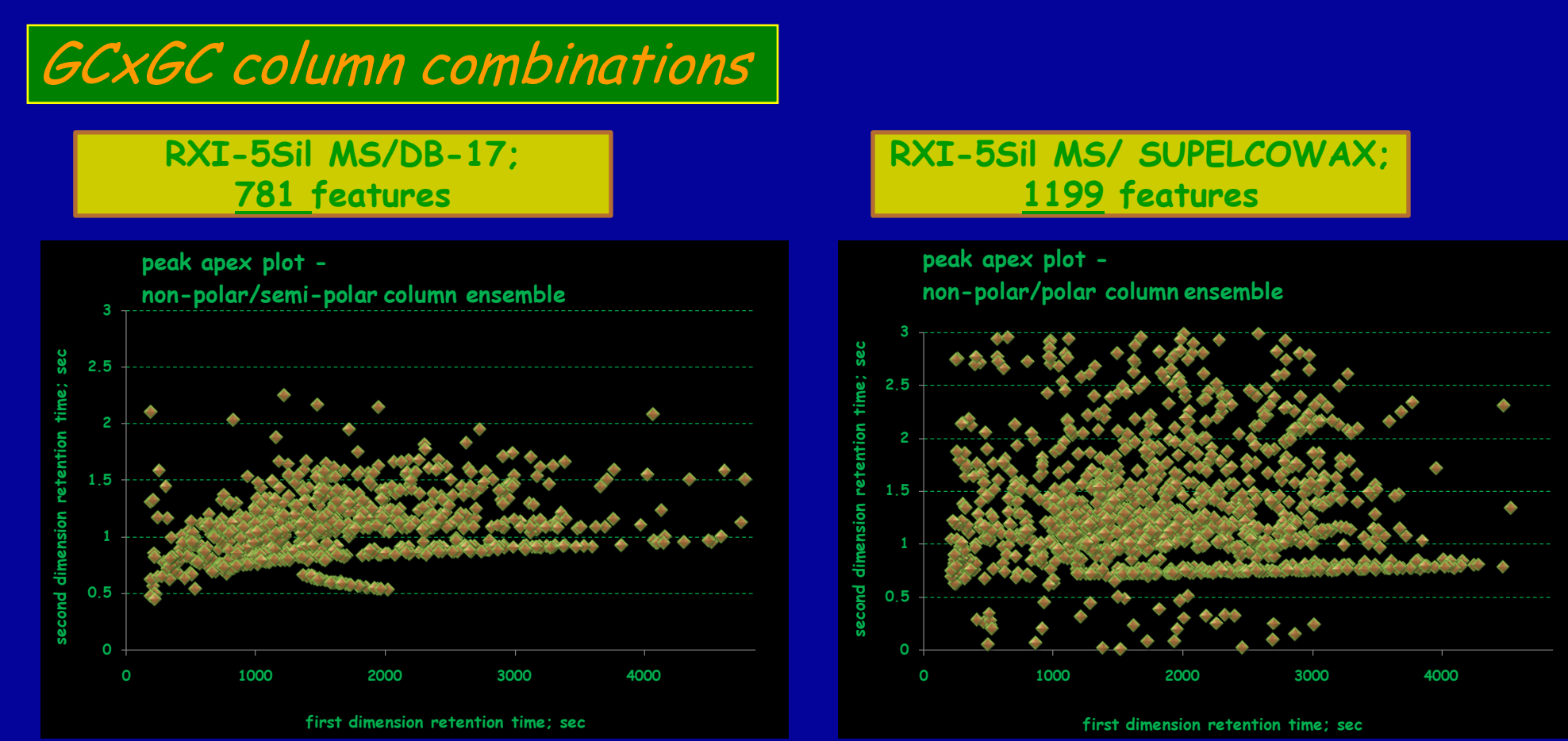
HS- & DI-SPME extraction modes of gaseous, liquid and solid samples;

↓ sample amount, ↓ sample preparation times & easy automation

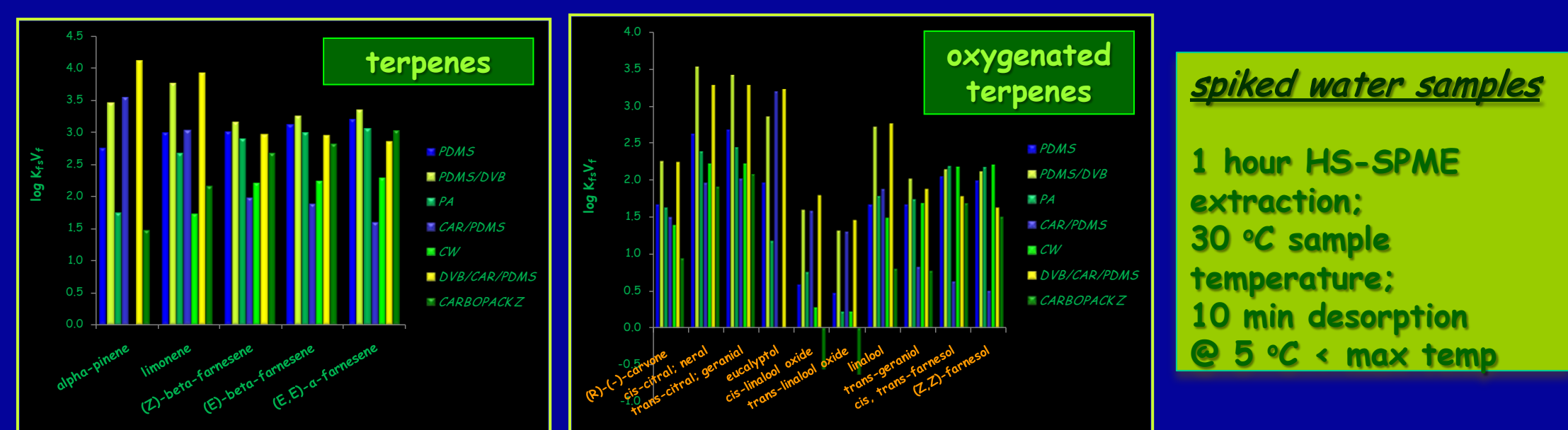
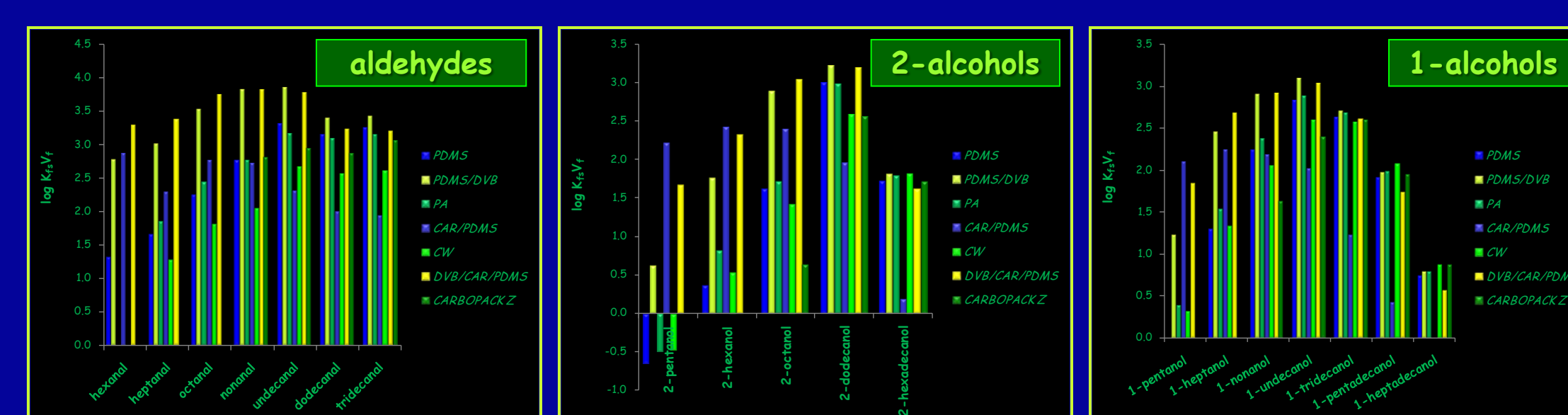
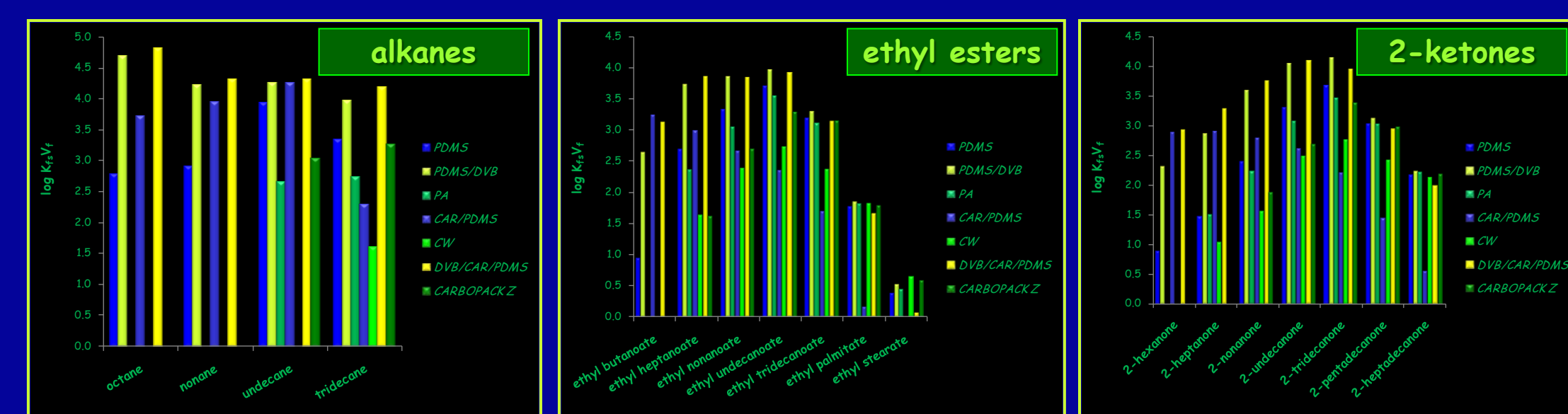
MINIATURIZED FORMAT & NON-EXHAUSTIVE EXTRACTION

↓ disturbance to investigated biological system;
 + *invivo* sampling: **metabolism quenching step eliminated**

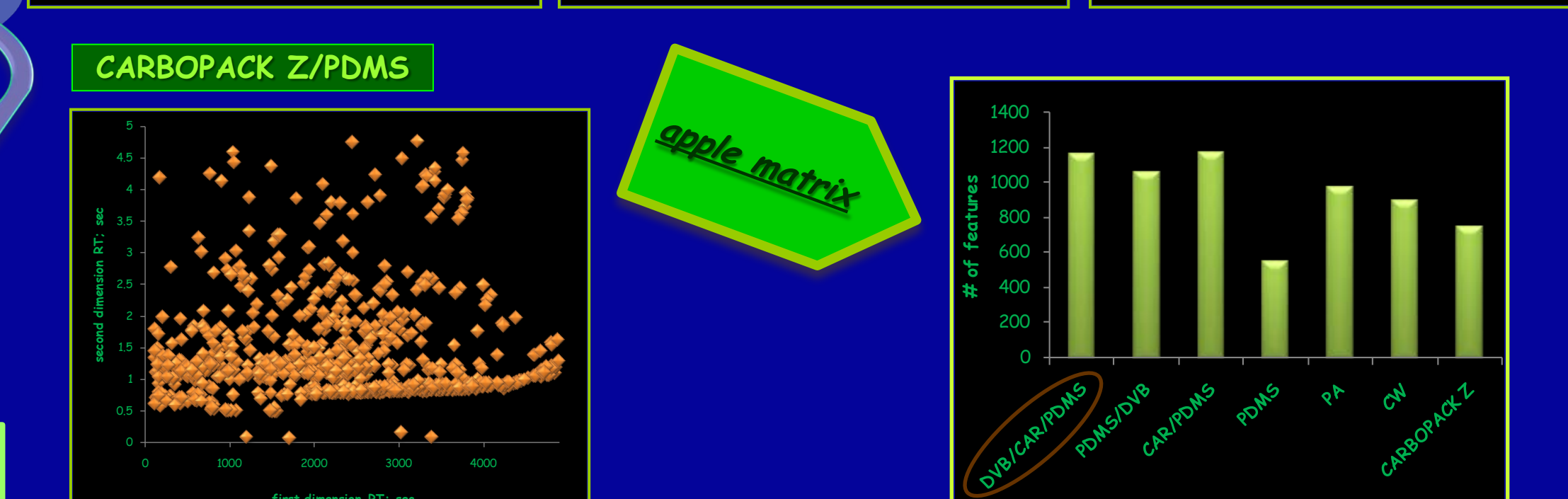
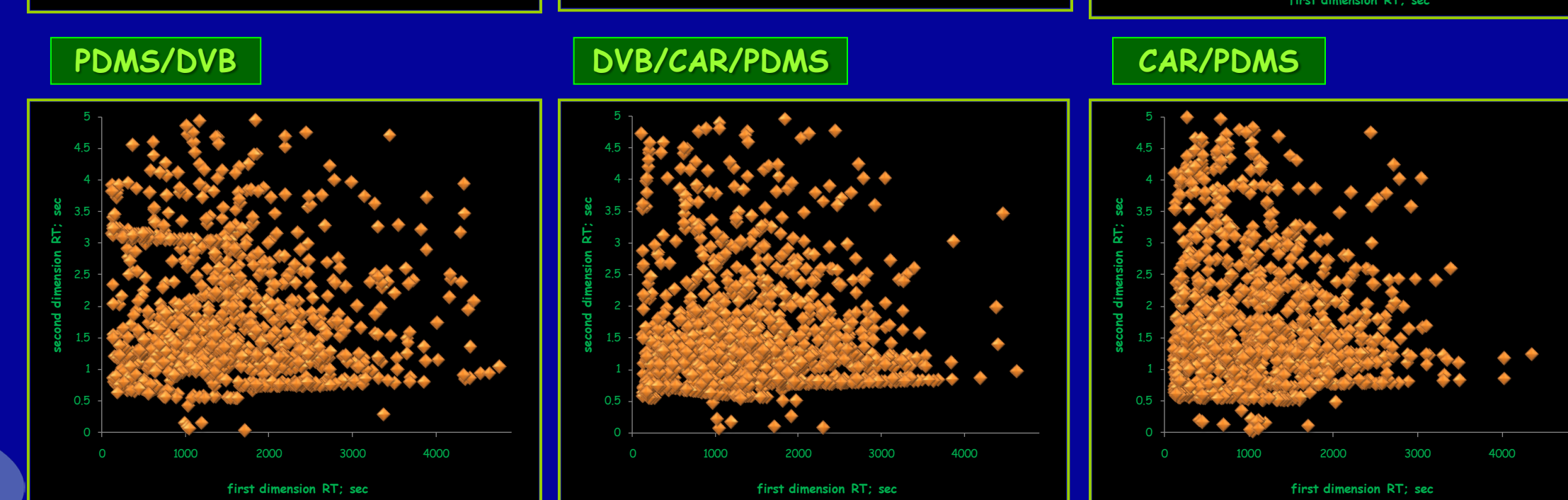
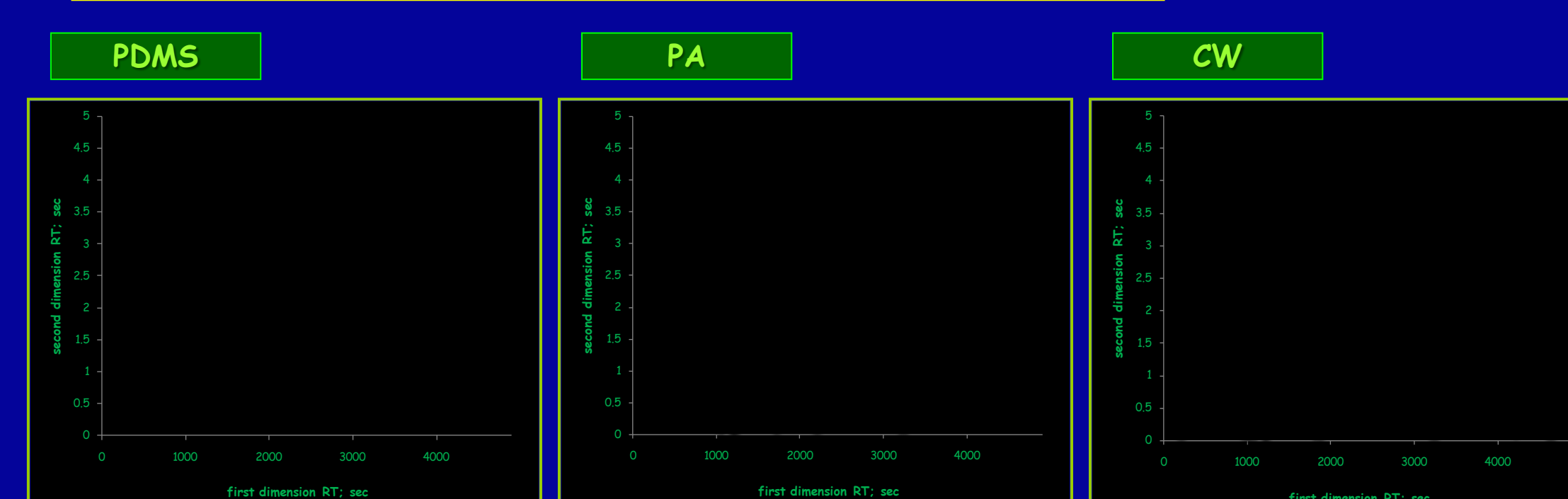
more representative sample extracts;
 more representative metabolism snapshot;
 ↓ sample preparation, extraction & storage artifacts;
 detection of rapid and short metabolism changes



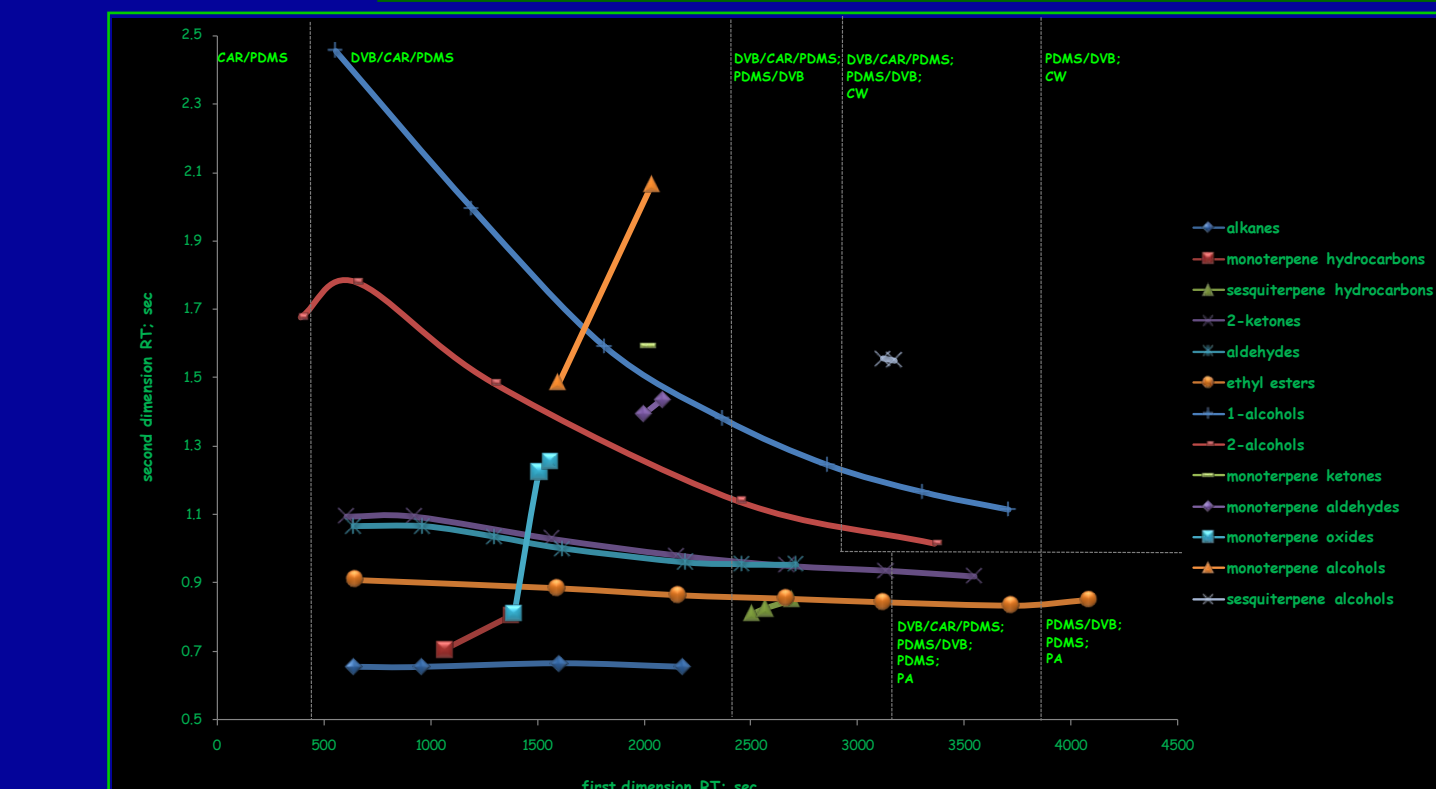
calculation of fibre coating/sample matrix distribution constants ($K_{f,s}$) & fibre constants ($K_{f,v}$)



SPME coating selection: extraction selectivity



toward standardization of SPME coating selection



MW & log K_{ow} thresholds:
 best sensitivity, selectivity & desorption efficiency

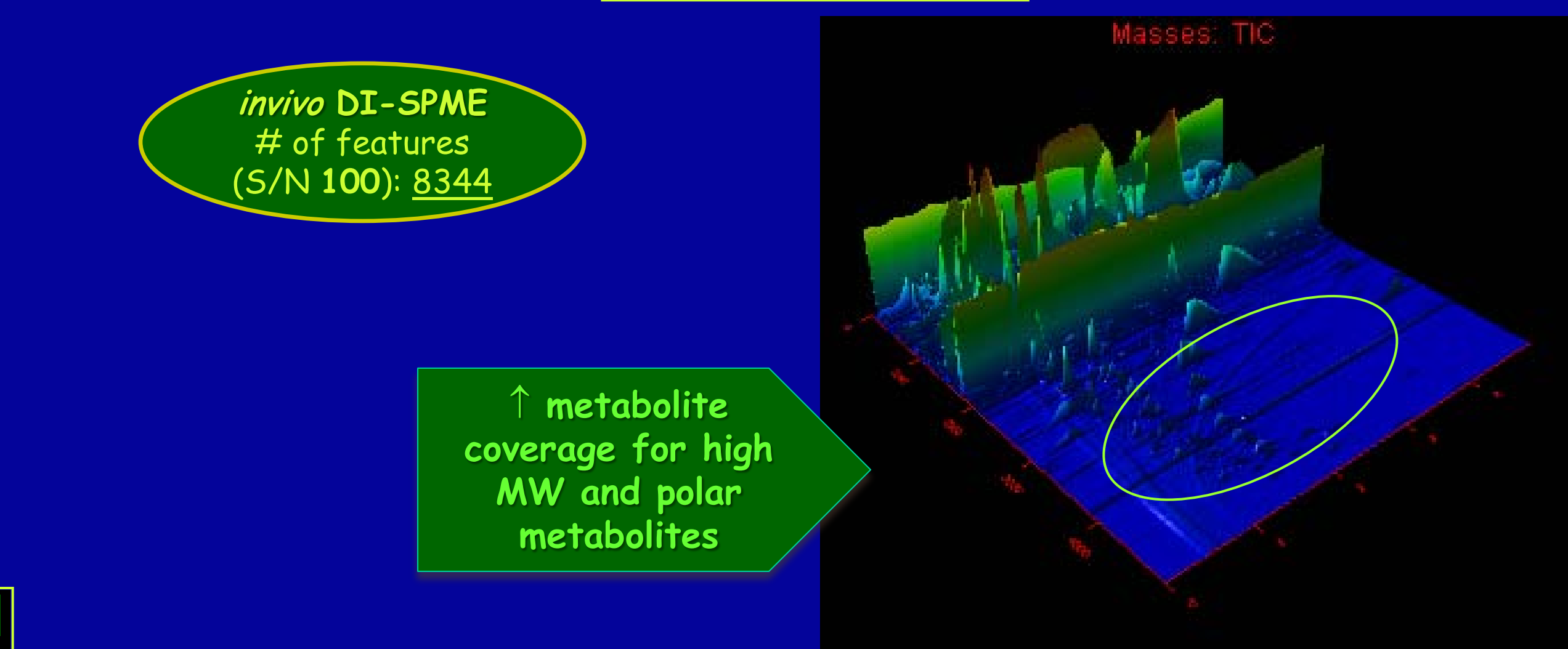
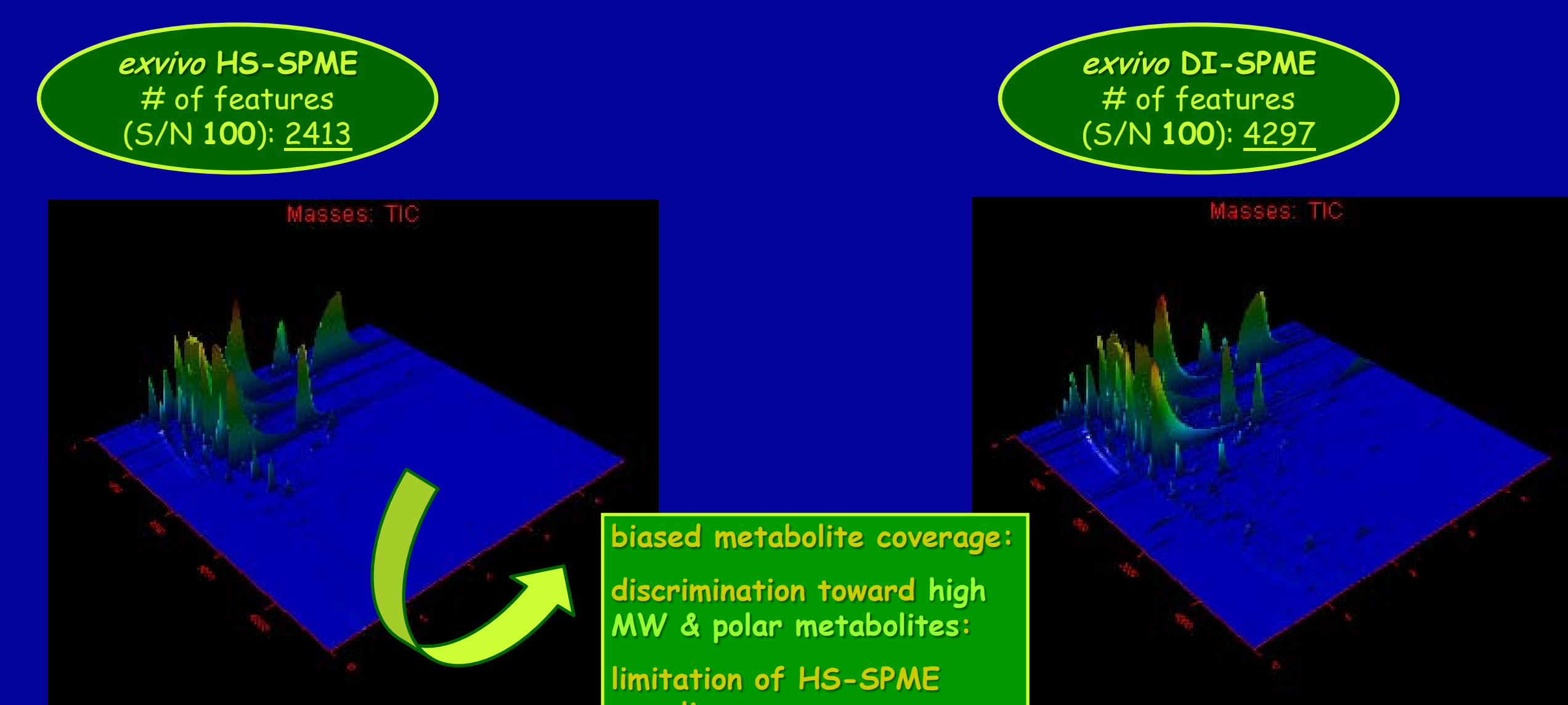
liquid sorbents

- non-polar compounds (alkanes and monoterpene hydrocarbons) slightly polar volatile compounds (ethyl esters, MW < 187 g/mol): PDMS
- more polar components (aliphatic aldehydes and ketones) sesquiterpene hydrocarbons; performance characteristics of PDMS and PA are not statistically different
- polar components having log K_{ow} < 3.30 (1-alcohols and 2-alcohols): CW and PA

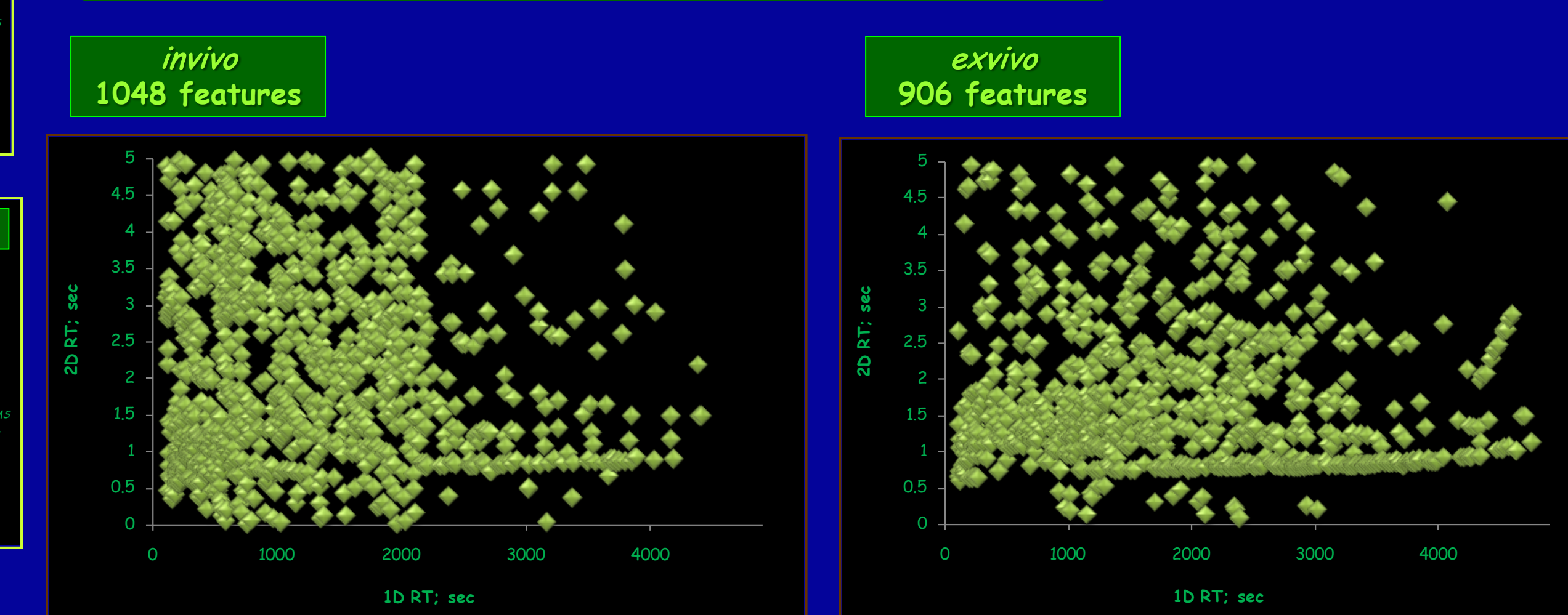
solid sorbents

- MW < 80-90 g/mol: CAR/PDMS
- MW 90-250 g/mol: DVB/CAR/PDMS
- MW 185-315 g/mol: PDMS/DVB

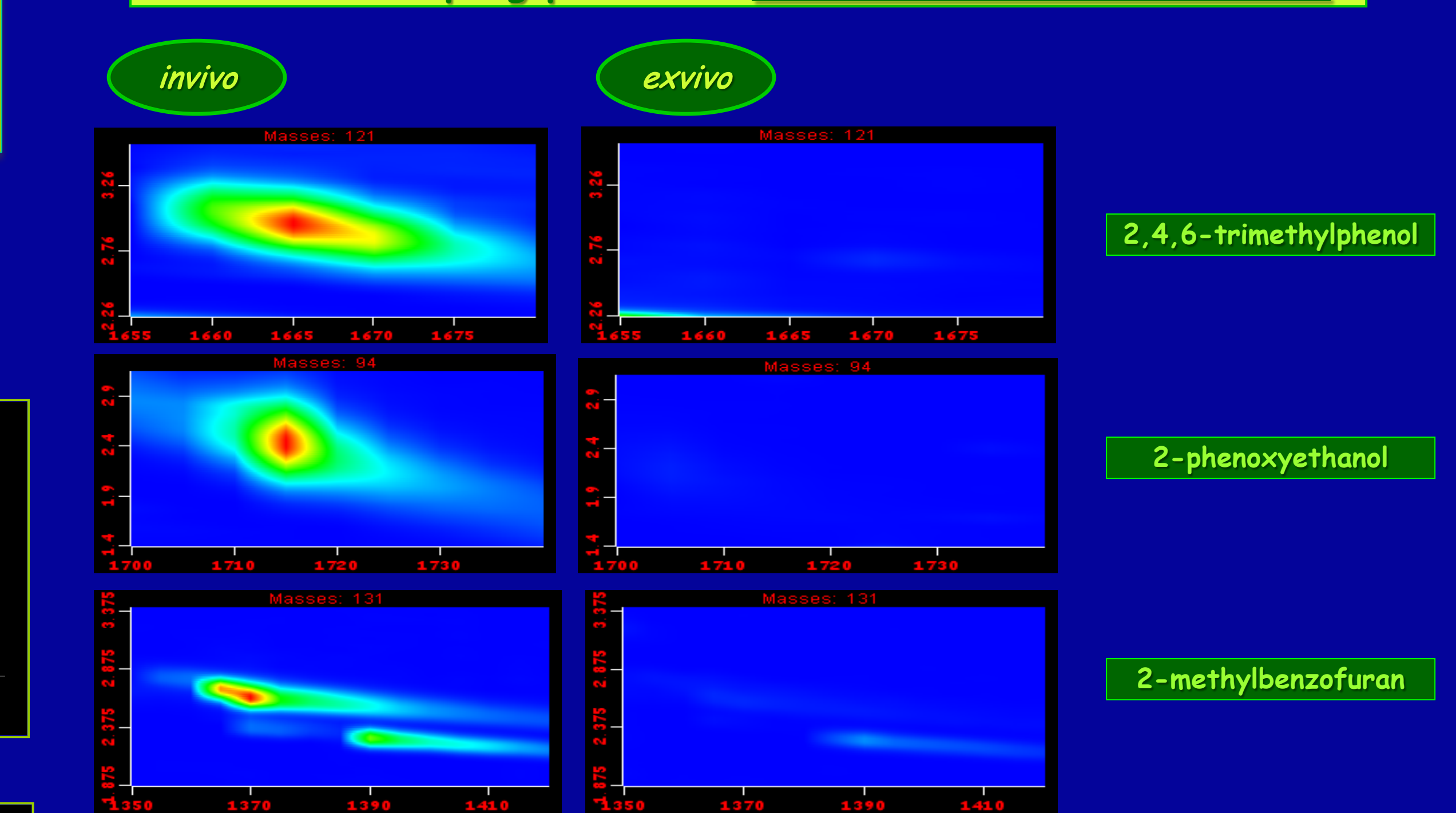
ex vivo & invivo sampling Δ in SPME sampling protocols: metabolite coverage



post-processing of peak tables for DI-SPME: S/N 200



Δ in SPME sampling protocols: differential metabolites invivo



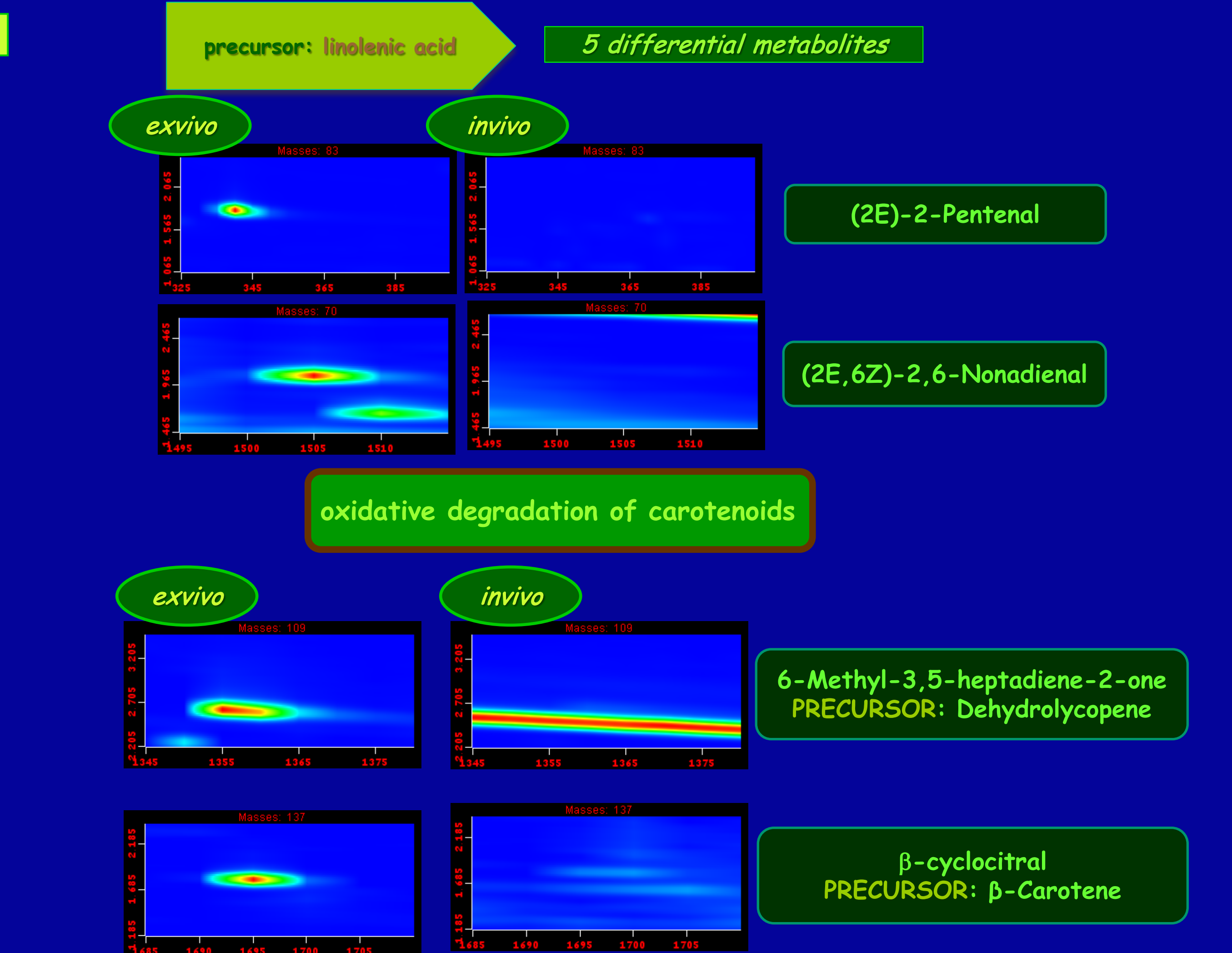
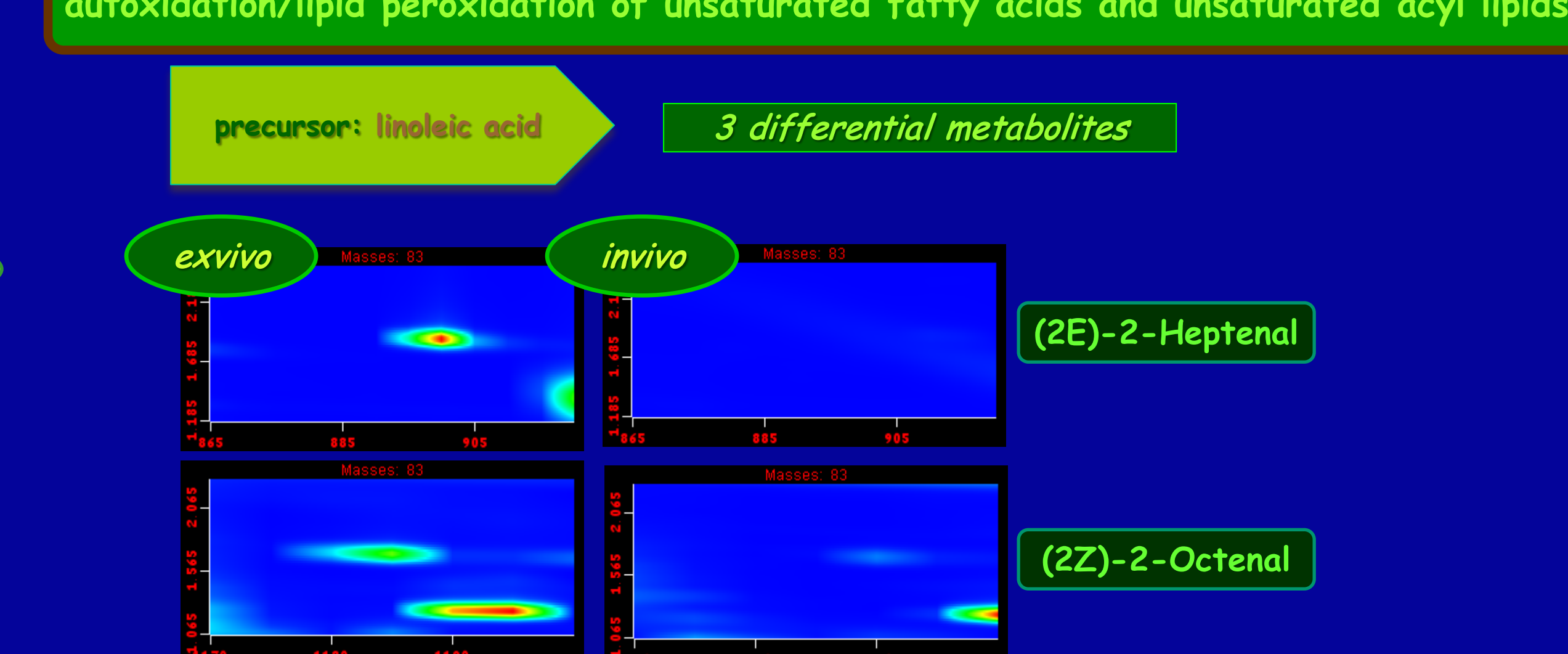
differential metabolites table

| analyte name | CAS # | RI _{inv} | RI _{ex} | similarity | structure |
|---------------------------|-----------|-------------------|------------------|------------|-----------|
| 2,4,6-Trimethylphenol | 527-60-6 | 1205 | 1204 | 824 | |
| 2-Phenoxyethanol | 122-99-6 | 1223 | 1226 | 898 | |
| gamma-Butyrolactone | 96-48-0 | 944 | 941 | 965 | |
| 2-Methylbenzofuran | 426-25-2 | 1105 | 1109 | 879 | |
| 2,2'-Bifuran | 5905-00-0 | 1037 | 1047 | 872 | |
| Butyl stearate | 123-95-5 | 2385 | 2374 | 862 | |
| 2-(2-Ethoxyethoxy)ethanol | 111-90-0 | 1019 | 1006 | 953 | |
| Z-11-Hexadecenoic acid | 2416-20-8 | 1938 | 1953 | 863 | |
| (2-Phenoxyethoxy)benzene | 104-66-5 | 1793 | 1811 | 892 | |

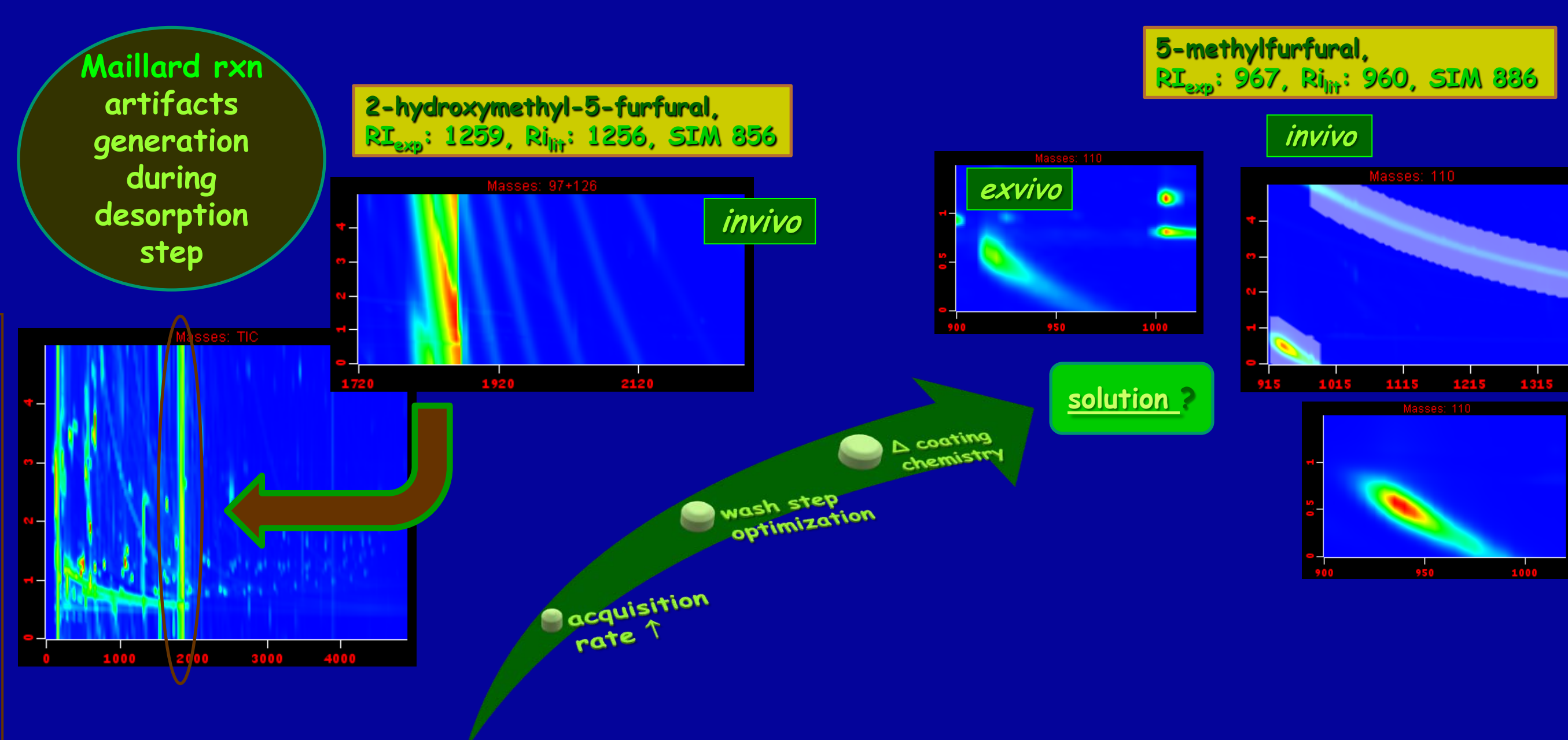
enzyme activation during ex vivo sample preparation step:
 non-specific degradation of metabolites?

Δ in SPME sampling protocols: differential metabolites ex vivo

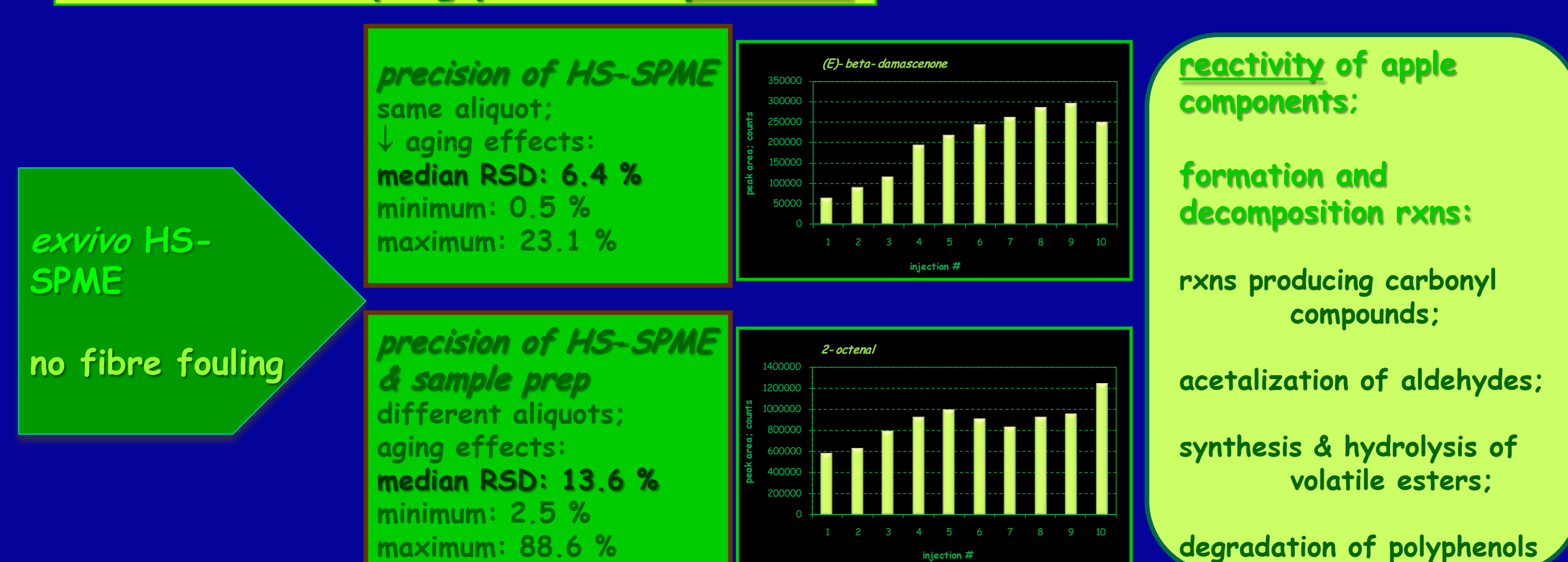
autooxidation/lipid peroxidation of unsaturated fatty acids and unsaturated acyl lipids



invivo sampling: CHALLENGES

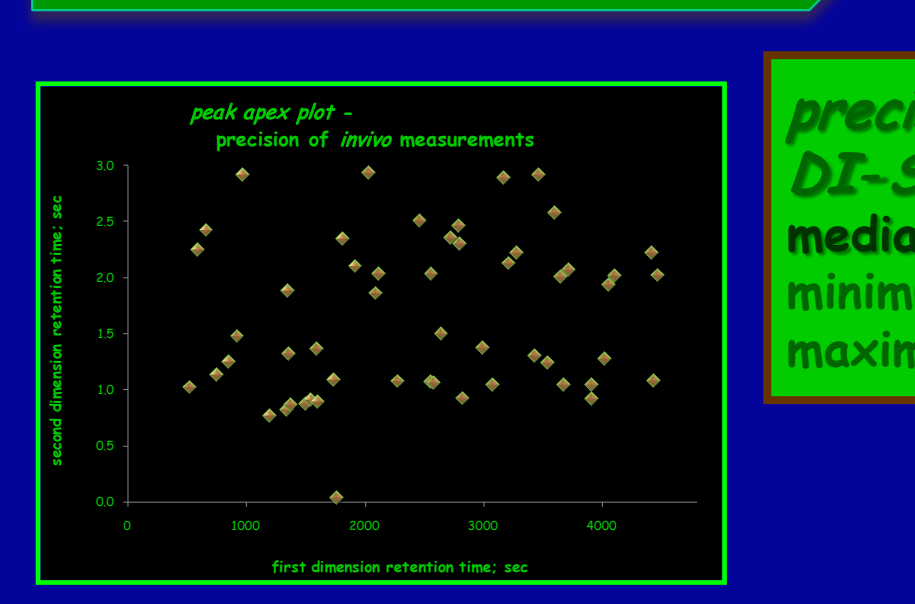


Δ in SPME sampling protocols: precision

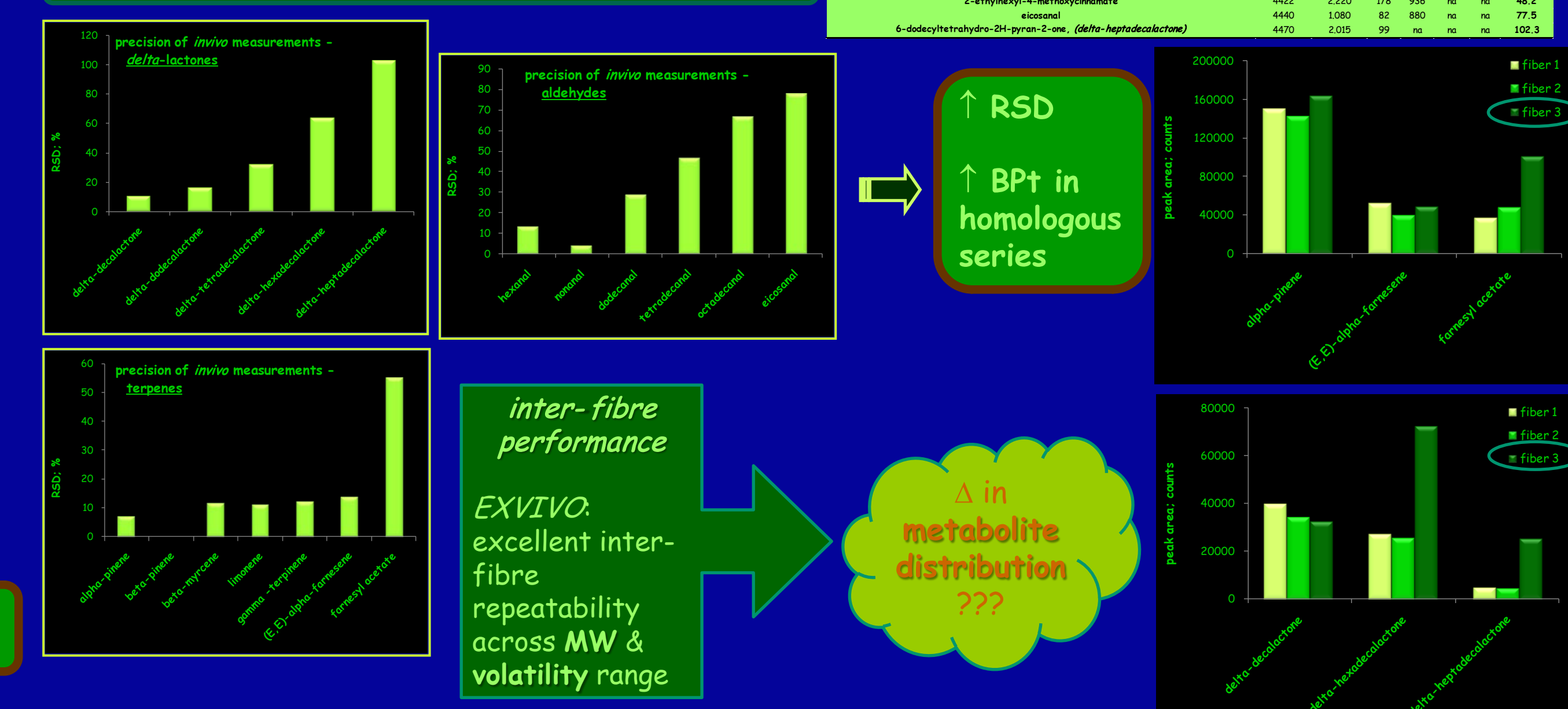


invivo DI-SPME

intra-fruit repeatability;
 n = 3 fibers, 53 compounds



close examination of selected functional groups



Project Objectives

SPME coating selection for GC-metabolomics

metabolite profiling of apples using SPME-GCxGC-TOFMS

comparison between *invivo* & *ex vivo* sampling formats

metabolite alignment, statistical analysis & identification of **biomarkers**

harvest & postharvest apple quality

Experimental

Pegasus 4D GCxGC-TOFMS system (LECO, St. Joseph, MI, USA):

Agilent 6890N GC high-speed TOF dual-stage quad-jet cryogenic modulator from Zoex & MPS 2 autosampler from Gerstel



SPME coating selection

comprehensive evaluation:
 extraction selectivity, extraction sensitivity & desorption efficiency of SPME coatings across a volatility & polarity range

MW range 88.15-312.54 g/mol;
 boiling point range 115.64-360.59 °C;
 log K_{ow} range 1.26-8.72

3 mL water sample spiked with 52 compounds belonging to homologous series

alkanes, alcohols, esters, aldehydes, monoterpenes, sesquiterpenes

GCxGC-TOFMS conditions: metabolomics samples

RXI-5 SIL MS x Supelcowax/BP20/Stabilwax columns,
 1.5 mL/min flow rate,
 40 °C (5 min), 3 °C/min to 240 °C (10 min),
 10 °C secondary oven offset,
 30 °C modulator temperature offset,
 5 sec modulation, 1 sec hot pulse time,
 m/z 33-950 acquisition range at 200 spectra/sec,
 1700 V detector voltage

SPME sampling protocols

- ex vivo** vs **invivo**
- metabolism quenching liquid nitrogen; saturated NaCl solution
 - homogenization
 - 1 hr HS-SPME & DI-SPME sampling
 - desorption & GCxGC-TOFMS analysis
- 1 hr DI-SPME sampling
 - wash step in water after sampling & before desorption
 - desorption & GCxGC-TOFMS analysis

Conclusions

potential of novel *invivo* SPME sampling formats and GCxGC-TOFMS for unbiased and representative characterization of metabolome
 ↑ metabolite coverage (1000s of metabolites at required sensitivities);
 median intra-fruit RSD - 16.3%;
 detection of differential metabolites

Acknowledgements

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